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(54) Title: A PROTEINASE INHIBITOR, PRECURSOR THEREOF AND GENETIC SEQUENCES ENCODING SAME			
(57) Abstract The present invention relates generally to proteinase inhibitors, a precursor thereof and to genetic sequences encoding same. More particularly, the present invention relates to a nucleic acid molecule comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a type II serin proteinase inhibitor (PI) precursor from a plant wherein said precursor comprises at least three PI monomers and wherein at least one of said monomers has a chymotrypsin specific site and at least one other of said monomers has a trypsin specific site.			

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A PROTEINASE INHIBITOR, PRECURSOR THEREOF AND GENETIC
SEQUENCES ENCODING SAME

- 5 The present invention relates generally to proteinase inhibitors, a precursor thereof and to genetic sequences encoding same.

Nucleotide and amino acid sequences are referred to herein by sequence identity numbers (SEQ ID NOs) which are defined after the bibliography. A general
10 summary of the SEQ ID NOs is provided before the examples.

Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated element or integer or group of
15 elements or integers but not the exclusion of any other element or integer or group of elements or integers.

Several members of the families Solanaceae and Fabaceae accumulate serine
proteinase inhibitors in their storage organs and in leaves in response to wounding
20 (Brown and Ryan, 1984; Richardson, 1977). The inhibitory activities of these proteins are directed against a wide range of proteinases of microbial and animal origin, but rarely against plant proteinases (Richardson, 1977). It is believed that these inhibitors are involved in protection of the plants against pathogens and predators. In potato tubers and legume seeds, the inhibitors can comprise 10% or
25 more of the stored proteins (Richardson, 1977), while in leaves of tomato and potato (Green and Ryan, 1972), and alfalfa (Brown and Ryan, 1984), proteinase inhibitors can accumulate to levels of 2% of the soluble protein within 48 hours of insect attack, or other types of wounding (Brown & Ryan, 1984; Graham *et al.*, 1986). High levels of these inhibitors (up to 50% of total soluble protein) are also present
30 in unripe fruits of the wild tomato, *Lycopersicon peruvianum* (Pearce *et al.*, 1988).

There are two families of serine proteinase inhibitors in tomato and potato (Ryan, 1984). Type I inhibitors are small proteins (monomer Mr 8100) which inhibit chymotrypsin at a single reactive site (Melville and Ryan, 1970; Plunkett *et al.*, 1982). Inhibitors of the type II family generally contain two reactive sites, one of which inhibits chymotrypsin and the other trypsin (Bryant *et al.*, 1976; Plunkett *et al.*, 1982). The type II inhibitors have a monomer Mr of 12,300 (Plunkett *et al.*, 1982). Proteinase inhibitor I accumulates in etiolated tobacco (*Nicotiana tabacum*) leaves (Kuo *et al.*, 1984), and elicitors from *Phytophthora parasitica* var. *nicotianae* were found to induce proteinase inhibitor I accumulation in tobacco cell suspension cultures (Rickauer *et al.*, 1989).

There is a need to identify other proteinase inhibitors and to investigate their potential use in the development of transgenic plants with enhanced protection against pathogens and predators. In accordance with the present invention, genetic sequences encoding a proteinase inhibitor precursor have been cloned. The precursor has multi-proteinase inhibitor domains and will be useful in developing a range of transgenic plants with enhanced proteinase inhibitor expression. Such plants will have enhanced protective properties against pathogens and predators. The genetic constructs of the present invention will also be useful in developing vaccines for ingestion by insects which are themselves predators or which act as hosts for plant pathogens. The recombinant precursor or monomeric inhibitors will also be useful in topical sprays and in assisting animals in feed digestion.

Accordingly, one aspect of the present invention relates to a nucleic acid molecule comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a type II serine proteinase inhibitor (PI) precursor from a plant wherein said precursor comprises at least three PI monomers and wherein at least one of said monomers has a chymotrypsin specific site and at least one other of said monomers has a trypsin specific site.

The "nucleic acid molecule" of the present invention may be RNA or DNA (eg cDNA), single or double stranded and linear or covalently closed. The nucleic acid molecule may also be genomic DNA corresponding to the entire gene or a substantial portion thereof or to fragments or derivatives thereof. The nucleotide sequence may correspond to the naturally occurring nucleotide sequence of the genomic or cDNA clone or may contain single or multiple nucleotide substitutions, deletions and/or additions thereto. All such variants in the nucleic acid molecule either retain the ability to encode at least one monomer or active part thereof or are useful as hybridisation probes or polymerase chain reaction (PCR) primers for the same or similar genetic sequences in other sources.

Preferably, the PI precursor comprises at least four, more preferably at least five and even more preferably at least six PI monomers. Still more preferably, the PI precursor further comprises a signal sequence. The PI precursor of the present invention preferably comprises amino acid sequences which are process sites for cleavage into individual monomers.

The term "precursor" as used herein is not intended to place any limitation on the utility of the precursor molecule itself or a requirement that the molecule first be processed into monomers before PI activity is expressed. The precursor molecule has PI activity and the present invention is directed to the precursor and to the individual monomers of the precursor.

Furthermore, the present invention extends to a nucleic acid molecule comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a hybrid type II serine PI precursor wherein said precursor comprises at least two monomers from different PIs. The at least two monomers may be modified such as being unable to be processed into individual monomers or may retain the ability to be so processed. Preferably, at least one of said monomers has a chymotrypsin specific site and the other of said monomers has a trypsin specific site. Preferably there are at least three monomers, more preferably at least four

- 4 -

monomers, still more preferably at least five monomers and yet still more preferably at least six monomers wherein at least two are from different PIs. In a most preferred embodiment, at least one of said monomers is a thionin. Such hybrid PI precursors and/or monomers thereof are particularly useful in generating molecules which are "multi-valent" in that they are active against a range of pathogens and predators such as both fungi and insects. Accordingly, reference herein to "PI precursor" includes reference to hybrid molecules.

The present invention is exemplified by the isolation of the subject nucleic acid molecule from *Nicotiana glauca* which has the following nucleotide sequence (SEQ ID NO. 1) and a corresponding amino acid sequence (SEQ ID NO. 3):

		AAG GCT TGT ACC TTA AAC
		Lys Ala Cys Thr Leu Asn
15		
	TGT GAT CCA AGA ATT GCC TAT GGA GTT TGC CCG CGT TCA GAA GAA AAG	
	Cys Asp Pro Arg Ile Ala Tyr Gly Val Cys Pro Arg Ser Glu Glu Lys	
	AAG AAT GAT CGG ATA TGC ACC AAC TGT TGC GCA GGC ACG AAG GGT TGT	
20	Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Thr Lys Gly Cys	
	AAG TAC TTC AGT GAT GAT GGA ACT TTT GTT TGT GAA GGA GAG TCT GAT	
	Lys Tyr Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp	
25	CCT AGA AAT CCA AAG GCT TGT ACC TTA AAC TGT GAT CCA AGA ATT GCC	
	Pro Arg Asn Pro Lys Ala Cys Thr Leu Asn Cys Asp Pro Arg Ile Ala	
	TAT GGA GTT TGC CCG CGT TCA GAA GAA AAG AAG AAT GAT CGG ATA TGC	
	Tyr Gly Val Cys Pro Arg Ser Glu Glu Lys Lys Asn Asp Arg Ile Cys	
30		
	ACC AAC TGT TGC GCA GGC ACG AAG GGT TGT AAG TAC TTC AGT GAT GAT	
	Thr Asn Cys Cys Ala Gly Thr Lys Gly Cys Lys Tyr Phe Ser Asp Asp	
	GGA ACT TTT GTT TGT GAA GGA GAG TCT GAT CCT AGA AAT CCA AAG GCT	
35	Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Arg Asn Pro Lys Ala	
	TGT CCT CGG AAT TGC GAT CCA AGA ATT GCC TAT GGG ATT TGC CCA CTT	
	Cys Pro Arg Asn Cys Asp Pro Arg Ile Ala Tyr Gly Ile Cys Pro Leu	

- 5 -

GCA GAA GAA AAG AAG AAT GAT CGG ATA TGC ACC AAC TGT TGC GCA GGC
Ala Glu Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly

5 AAA AAG GGT TGT AAG TAC TTT AGT GAT GAT GGA ACT TTT GTT TGT GAA
Lys Lys Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr Phe Val Cys Glu

GGA GAG TCT GAT CCT AAA AAT CCA AAG GCC TGT CCT CGG AAT TGT GAT
Gly Glu Ser Asp Pro Lys Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp

10 GGA AGA ATT GCC TAT GGG ATT TGC CCA CTT TCA GAA GAA AAG AAG AAT
Gly Arg Ile Ala Tyr Gly Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn

GAT CGG ATA TGC ACC AAC TGC TGC GCA GGC AAA AAG GGT TGT AAG TAC
15 Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr

TTT AGT GAT GAT GGA ACT TTT GTT TGT GAA GGA GAG TCT GAT CCT AAA
Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Lys

20 AAT CCA AAG GCT TGT CCT CGG AAT TGT GAT GGA AGA ATT GCC TAT GGG
Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly

ATT TGC CCA CTT TCA GAA GAA AAG AAG AAT GAT CGG ATA TGC ACA AAC
Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn

25 TGT TGC GCA GGC AAA AAG GGC TGT AAG TAC TTT AGT GAT GAT GGA ACT
Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr

TTT GTT TGT GAA GGA GAG TCT GAT CCT AGA AAT CCA AAG GCC TGT CCT
30 Phe Val Cys Glu Gly Glu Ser Asp Pro Arg Asn Pro Lys Ala Cys Pro

CGG AAT TGT GAT GGA AGA ATT GCC TAT GGA ATT TGC CCA CTT TCA GAA
Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly Ile Cys Pro Leu Ser Glu

35 GAA AAG AAG AAT GAT CGG ATA TGC ACC AAT TGT TGC GCA GGC AAG AAG
Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys

GGC TGT AAG TAC TTT AGT GAT GAT GGA ACT TTT ATT TGT GAA GGA GAA
Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr Phe Ile Cys Glu Gly Glu

40 TCT GAA TAT GCC AGC AAA GTG GAT GAA TAT GTT GGT GAA GTG GAG AAT
Ser Glu Tyr Ala Ser Lys Val Asp Glu Tyr Val Gly Glu Val Glu Asn

GAT CTC CAG AAG TCT AAG GTT GCT GTT TCC
45 Asp Leu Gln Lys Ser Lys Val Ala Val Ser

- 6 -

This is done, however, with the understanding that the present invention extends to an equivalent or substantially similar nucleic acid molecule from any other plant. By "equivalent" and "substantially similar" is meant at the level of nucleotide sequence, amino acid sequence, antibody reactivity, monomer composition and/or processing of the precursor to produce monomers. For example, a nucleotide sequence having a percentage sequence similarity of at least 55%, such as about 60-65%, 70-75%, 80-85% and over 90% when compared to the sequence of SEQ ID NO. 1 would be considered "substantially similar" to the subject nucleic acid molecule provided that such a substantially similar sequence encodes a PI precursor having at least three monomers and preferably four, five or six monomers as hereinbefore described.

In a particularly preferred embodiment, the nucleic acid molecule further encodes a signal sequence 5' to the open reading frame and/or a nucleotide sequence 3' of the coding region providing a full nucleotide sequence as follows (SEQ ID NO. 2):

```
CGAGTAAGTA TGGCTGTTCA CAGAGTTAGT TTCCTTGCTC TCCTCCTCTT ATTTGGAATG

TCTCTGCTTG TAAGCAATGT GGAACATGCA GATGCC AAG GCT TGT ACC TTA AAC
20                               Lys Ala Cys Thr Leu Asn

TGT GAT CCA AGA ATT GCC TAT GGA GTT TGC CCG CGT TCA GAA GAA AAG
Cys Asp Pro Arg Ile Ala Tyr Gly Val Cys Pro Arg Ser Glu Glu Lys

25 AAG AAT GAT CGG ATA TGC ACC AAC TGT TGC GCA GGC ACG AAG GGT TGT
Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Thr Lys Gly Cys

AAG TAC TTC AGT GAT GAT GGA ACT TTT GTT TGT GAA GGA GAG TCT GAT
Lys Tyr Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp
30

CCT AGA AAT CCA AAG GCT TGT ACC TTA AAC TGT GAT CCA AGA ATT GCC
Pro Arg Asn Pro Lys Ala Cys Thr Leu Asn Cys Asp Pro Arg Ile Ala

TAT GGA GTT TGC CCG CGT TCA GAA GAA AAG AAG AAT GAT CGG ATA TGC
35 Tyr Gly Val Cys Pro Arg Ser Glu Glu Lys Lys Asn Asp Arg Ile Cys

ACC AAC TGT TGC GCA GGC ACG AAG GGT TGT AAG TAC TTC AGT GAT GAT
Thr Asn Cys Cys Ala Gly Thr Lys Gly Cys Lys Tyr Phe Ser Asp Asp
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- 7 -

GGA ACT TTT GTT TGT GAA GGA GAG TCT GAT CCT AGA AAT CCA AAG GCT
Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Arg Asn Pro Lys Ala

5 TGT CCT CGG AAT TGC GAT CCA AGA ATT GCC TAT GGG ATT TGC CCA CTT
Cys Pro Arg Asn Cys Asp Pro Arg Ile Ala Tyr Gly Ile Cys Pro Leu

GCA GAA GAA AAG AAG AAT GAT CGG ATA TGC ACC AAC TGT TGC GCA GGC
Ala Glu Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly

10 AAA AAG GGT TGT AAG TAC TTT AGT GAT GAT GGA ACT TTT GTT TGT GAA
Lys Lys Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr Phe Val Cys Glu

GGA GAG TCT GAT CCT AAA AAT CCA AAG GCC TGT CCT CGG AAT TGT GAT
Gly Glu Ser Asp Pro Lys Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp

15 GGA AGA ATT GCC TAT GGG ATT TGC CCA CTT TCA GAA GAA AAG AAG AAT
Gly Arg Ile Ala Tyr Gly Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn

GAT CGG ATA TGC ACC AAC TGC TGC GCA GGC AAA AAG GGT TGT AAG TAC
20 Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr

TTT AGT GAT GAT GGA ACT TTT GTT TGT GAA GGA GAG TCT GAT CCT AAA
Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Lys

25 AAT CCA AAG GCT TGT CCT CGG AAT TGT GAT GGA AGA ATT GCC TAT GGG
Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly

ATT TGC CCA CTT TCA GAA GAA AAG AAG AAT GAT CGG ATA TGC ACA AAC
Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn

30 TGT TGC GCA GGC AAA AAG GGC TGT AAG TAC TTT AGT GAT GAT GGA ACT
Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr

TTT GTT TGT GAA GGA GAG TCT GAT CCT AGA AAT CCA AAG GCC TGT CCT
35 Phe Val Cys Glu Gly Glu Ser Asp Pro Arg Asn Pro Lys Ala Cys Pro

CGG AAT TGT GAT GGA AGA ATT GCC TAT GGA ATT TGC CCA CTT TCA GAA
Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly Ile Cys Pro Leu Ser Glu

40 GAA AAG AAG AAT GAT CGG ATA TGC ACC AAT TGT TGC GCA GGC AAG AAG
Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys

GGC TGT AAG TAC TTT AGT GAT GAT GGA ACT TTT ATT TGT GAA GGA GAA
Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr Phe Ile Cys Glu Gly Glu

45

- 8 -

TCT GAA TAT GCC AGC AAA GTG GAT GAA TAT GTT GGT GAA GTG GAG AAT
Ser Glu Tyr Ala Ser Lys Val Asp Glu Tyr Val Gly Glu Val Glu Asn

GAT CTC CAG AAG TCT AAG GTT GCT GTT TCC TAAGTCCTAA CTAATAATAT
5 Asp Leu Gln Lys Ser Lys Val Ala Val Ser

GTAGTCTATG TATGAAACAA AGGCATGCCA ATATGCTCTG TCTTGCCTGT AATCTGTAAT

ATGGTAGTGG AGCTTTTCCA CTGCCTGTTT AATAAGAAAT GGAGCACTAG TTTGTTTTAG
10 TTAAAAAAAA AAAAAAAAAA

including substantially similar variants thereof.

15 Accordingly, a preferred embodiment of the present invention provides a nucleic acid molecule comprising a sequence of nucleotides as set forth in SEQ ID NO. 1 or 2 which encodes or is complementary to a sequence which encodes a type II serine PI precursor from *Nicotiana alata* or having at least 55% similarity to said precursor or at least one domain therein wherein said precursor comprises a signal peptide and
20 at least five monomers and wherein one of said monomers has a chymotrypsin specific site and four of said monomers have trypsin specific sites.

In still a more preferred embodiment, the nucleic acid molecule is a cDNA molecule and comprises a nucleotide sequence generally as set forth in SEQ ID NO. 1 or 2
25 or being substantially similar thereto as hereinbefore defined to the whole of said sequence or to a domain thereof.

Another aspect of the present invention is directed to a nucleic acid molecule comprising a sequence of nucleotides which encodes or is complementary to a
30 sequence which encodes a single type II serine PI having either a chymotrypsin specific site or a trypsin specific site and wherein said PI is a monomer of a precursor PI having at least three monomers of which at least one of said monomers has a chymotrypsin site and the other of said monomers has a trypsin site. Preferably, however, the precursor has four, five or six monomers and is as hereinbefore defined.

35

In its most preferred embodiment, the plant is *N. alata* (Link et Otto) having self-incompatibility genotype S_1S_3 , S_3S_3 or S_6S_6 , and the nucleic acid molecule is isolatable from or complementary to genetic sequences isolatable from stigmas and styles of mature plants. The corresponding mRNA is approximately 1.4 kb and the cDNA has six conserved domains wherein the first two domains are 100% identical and contain chymotrypsin-specific sites (Leu-Asn). The third, fourth and fifth domains share 95-98% identity and have sites specific for trypsin (Arg-Asn). A sixth domain which also has a trypsin specific site has less identity to the third, fourth and fifth domains (79-90%) due mainly to a divergent 3' sequence (see Table 1). The preferred PI inhibitor of the present invention has a molecular weight of approximately 42-45 kDa with an approximately 29 amino acid signal sequence.

The N-terminal sequence of the monomeric PI is represented in each of the six repeated domains in the predicted sequence of the PI precursor protein. Thus, it is likely that the PI precursor protein is cleaved at six sites to produce seven peptides. Six of the seven peptides, peptides 2, 3, 4, 5, 6 and 7 (Figure 1, residues 25-82 [SEQ ID NO. 5], 83-140 [SEQ ID NO. 6], 141-198 [SEQ ID NO. 7], 199-256 [SEQ ID NO. 8], 257-314 [SEQ ID NO. 9] and 315-368 [SEQ ID NO. 9], respectively), would be in the same molecular weight range as the monomeric PI (about 6 kDa) and would have the same N-terminal sequence. Peptide 7 does not contain a consensus site for trypsin or chymotrypsin. Peptide 1 (residues 1-24 [SEQ ID NO. 4], Figure 1) is smaller than 6 kD, has a different N-terminus and was not detected in a purified monomeric PI preparation. It could be envisaged that peptide 1 and peptide 7 would form a functional proteinase inhibitor with the inhibitory site on peptide 1 held in the correct conformation by disulphide bonds formed between the two peptides.

Although not intending to limit the present invention to any one hypothesis, the PI precursor may be processed by a protease responsible, for example, for cleavage of an Asn-Asp linkage, to produce the bioactive monomers. More particularly, the protease sensitive sequence is $R_1-X_1-X_2$ -Asn-Asp- R_2 where R_1 , R_2 , X_1 and X_2 are

- 10 -

defined below. The discovery of such a sequence will enable the engineering of peptides and polypeptides capable of being processed in a plant by cleavage of the protease sensitive sequence. According to this aspect of the present invention there is provided a protease sensitive peptide comprising the amino acid sequence:

5



wherein X_1 and X_2 are any amino acid but are preferably both Lys residues. The protease sensitive peptide may also be represented as:

10



wherein X_1 and X_2 are preferably the same and are preferably both Lys residues and wherein R_1 and R_2 are the same or different, any D or L amino acid, a peptide, a polypeptide, a protein, or a non-amino acid moiety or molecule such as, but not limited to, an alkyl (eg methyl, ethyl), substituted alkyl, alkenyl, substituted alkenyl, acyl, dienyl, arylalkyl, arylalkenyl, aryl, substituted aryl, heterocyclic, substituted heterocyclic, cycloalkyl, substituted cycloalkyl, halo (e.g. Cl, Br, I, F), haloalkyl, nitro, hydroxy, thiol, sulfonyl, carboxy, alkoxy, aryloxy and alkylaryloxy group and the like as would be apparent to one skilled in the art. By alkyl, substituted alkyl, alkenyl and substituted alkenyl and the like is meant to encompass straight and branched molecules, lower ($C_1 - C_6$) and higher (more than C_6) derivatives. The term "substituted" includes all the substituents set forth above.

25 In its most preferred embodiment, the protease sensitive peptide is:



wherein R_1 and R_2 are the same or different and are peptides or polypeptides and
30 wherein X_1 and X_2 are both Lys residues.

Such a protease sensitive peptide can be placed between the same or different monomers so that upon expression in a suitable host or *in vitro*, the larger molecule can be processed to the peptides located between the protease sensitive peptides.

5

The present invention also extends to a nucleic acid molecule comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a protease sensitive peptide comprising the sequence:

10



wherein X_1 and X_2 are preferably the same and are most preferably both Lys residues. Such a nucleic acid molecule may be part of a larger nucleotide sequence encoding, for example, a precursor polypeptide capable of being processed via the protease sensitive sequence into individual peptides or monomers.

15

The protease sensitive peptide of the present invention is particularly useful in generating poly and/or multi-valent "precursors" wherein each monomer is the same or different and directed to the same or different activities such as anti-viral, anti-bacterial, anti-fungal, anti-pathogen and/or anti-predator activity.

20

Although not wishing to limit this aspect of the invention to any one hypothesis or proposed mechanism of action, it is believed that the protease acts adjacent the Asn residue as more particularly between the Asn-Asp residues.

25

The present invention extends to an isolated type II serine PI precursor from a plant wherein said precursor comprises at least three PI monomers and wherein at least one of said monomers has a chymotrypsin specific site and at least one other of said monomers has a trypsin specific site. Preferably, the PI precursor has four, five or six monomers and is encoded by the nucleic acid molecule as hereinbefore described. The present invention also extends to the individual monomers comprising the precursor. The present invention also extends to a hybrid recombinant PI precursor

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- 12 -

molecule comprising at least two monomers from different PIs as hereinbefore described.

The isolated PI or PI precursor may be in recombinant form and/or biologically
5 pure. By "biologically pure" is meant a preparation of PI, PI precursor and/or any
mixtures thereof having undergone at least one purification step including
ammonium sulphate precipitation, Sephadex chromatography and/or affinity
chromatography. Preferably, the preparation comprises at least 20% of the PI, PI
precursor or mixture thereof as determined by weight, activity antibody, reactivity
10 and/or amino acid content. Even more preferably, the preparation comprises 30-
40%, 50-60% or at least 80-90% of PI, PI precursor or mixture thereof.

The PI or its precursor may be naturally occurring or be a variant as encoded by the
nucleic acid variants referred to above. It may also contain single or multiple
15 substitutions, deletions and/or additions to its amino acid sequence or to non-
proteinaceous components such as carbohydrate and/or lipid moieties.

The recombinant and isolated PI, PI precursor and mixtures thereof are useful as
laboratory reagents, in the generation of antibodies, in topically applied insecticides
20 as well as orally ingested insecticides.

The recombinant PI or PI precursor may be provided as an insecticide alone or in
combination with one or more carriers or other insecticides such as the BT crystal
protein.
25

The PI of the present invention is considered to have a defensive role in organs of
the plant, for example, the stigma, against the growth or infection by pests and
pathogens such as fungi, bacteria and insects. There is a need, therefore, to develop
genetic constructs which can be used to generate transgenic plants capable of
30 expressing the PI precursor where this can be processed into monomers of a
monomeric PI itself.

- 13 -

Accordingly, another aspect of the present invention contemplates a genetic construct comprising a nucleic acid molecule comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a type II serine PI precursor or monomer thereof from a plant wherein said precursor comprises at least
5 three PI monomers and wherein at least one of said monomers has a chymotrypsin specific site and at least one of said other monomers has a trypsin specific site and said genetic sequence further comprises expression means to permit expression of said nucleic acid molecule, replication means to permit replication in a plant cell or, alternatively, integration means, to permit stable integration of said nucleic acid
10 molecule into a plant cell genome. Preferably, the expression is regulated such as developmentally or in response to infection such as being regulated by an existing PI regulatory sequence. Preferably, the expression of the nucleic acid molecule is enhanced to thereby provide greater endogenous levels of PI relative to the levels in the naturally occurring plant. Alternatively, the PI precursor cDNA of the present
15 invention is usable to obtain a promoter sequence which can then be used in the genetic construct or to cause its manipulation to thereby permit over-expression of the equivalent endogenous promoter. In another embodiment the PI precursor is a hybrid molecule as hereinbefore described.

20 Yet another aspect of the present invention is directed to a transgenic plant carrying the genetic sequence and/or nucleic acid molecule as hereinbefore described and capable of producing elevated, enhanced or more rapidly produced levels of PI and/or PI precursor or hybrid PI precursor when required. Preferably, the plant is a crop plant or a tobacco plant but other plants are usable where the PI or PI
25 precursor nucleic acid molecule is expressable in said plant. Where the transgenic plant produces PI precursor, the plant may or may not further process the precursor into monomers. Alternatively, the genetic sequence may be part of a viral or bacterial vector for transmission to an insect to thereby control pathogens in insects which would consequently interrupt the transmission of the pathogens to plants.

30

In still yet another aspect of the present invention, there is provided antibodies to the PI precursor or one or more of its monomers. Antibodies may be monoclonal or polyclonal and are useful in screening for PI or PI precursor clones in an expression library or for purifying PI or PI precursor in a fermentation fluid, supernatant fluid or plant extract.

The genetic constructs of the present invention can also be used to populate the gut of insects to act against the insect itself or any plant pathogens therein or to incorporate into the gut of animals to facilitate the digestion of plant material.

The present invention is further described by reference to the following non-limiting Figures and Examples.

15 In the Figures:

Figure 1 shows the nucleic acid sequence (SEQ ID NO. 2) of the pNA-PI-2 insert and the corresponding amino acid sequence (SEQ ID NO. 3) of the *N. alata* PI protein. The amino acid sequence is numbered beginning with 1 for the first amino acid of the mature protein. The signal sequence is encoded by nucleotides 1 to 97 and the amino acid residues have been assigned negative numbers. The reactive site residues of the inhibitor are boxed. The *N. alata* PI sequence contains six similar domains (domain 1, residues 1 to 58, domain 2, residues 59-116, domain 3, residues 117-174, domain 4, residues 175-232, domain 5, residues 233-290 and domain 6, residues 291-343).

Figure 2 is a photographic representation showing a gel blot analysis of RNA from various organs of *N. alata*. Gel Blot of RNA isolated from organs of *N. alata* and from stigmas and styles of *N. tabacum* and *N. sylvestris*, hybridised with the cDNA clone NA-PI-2. St, stigma and style; Ov, ovaries; Po, pollen; Pe, petals; Se, sepals; L, non-wounded leaves; L4, leaves 4h after wounding; L24, leaves 24h after wounding; Nt, *N. tabacum* stigma and style; Ns, *N. sylvestris* stigma and style; Na

- 15 -

*Hind*III restriction fragments of Lambda-DNA.

The NA-PI-2 clone hybridised to 2 mRNA species (1.0 and 1.4kb). The larger mRNA was predominant in stigma and styles, whereas the smaller mRNA species was more dominant in other tissues. After high stringency washes, the 1.0kb mRNA from stigma and style no longer hybridises to the NA-PI-2 probe.

Figure 3 is a photographic presentation depicting *in situ* localisation of RNA homologous to NA-PI-2 in stigma and style.

- 10 (a) Autoradiograph of a longitudinal cryosection through the stigma and style of a 1cm long bud after hybridisation with the ³²P-labelled NA-PI-2 cDNA probe.
- (b) The same section as (a), stained with toluidine blue. c, cortex; v, vascular bundles; tt, transmitting tract; s, stigmatic tissue.

15

The cDNA probe labelled the cells of the stigma heavily and some hybridisation to the vascular bundles can be seen. There was no hybridisation to the epidermis, cortical tissue or transmitting tissue. Scale bars = 200 μ m.

- 20 Figure 4 is a photographic representation of a gel blot analysis of genomic DNA of *N. alata*. Gel blot analysis of *N. alata* genomic DNA digested with the restriction enzymes *Eco*RI or *Hind*III, and probed with radiolabelled NA-PI-2. Size markers (kb) are *Hind*III restriction fragments of Lambda-DNA.

- 25 *Eco*RI produced two hybridising fragments (11kb and 7.8kb), while *Hind*III gave three large hybridising fragments (16.6, 13.5 and 10.5kb). The NA-PI-2 clone appears to belong to a small multigene family consisting of at least two members.

- Figure 5 is a graphic representation of PI activity in various organs of *N. alata*.
- 30 Buffer soluble extracts from various organs were tested for their ability to inhibit trypsin and chymotrypsin. Stigma and sepal extracts were the most effective inhibitors of both trypsin (A) and chymotrypsin (B).

- 16 -

Figure 6 depicts the steps of the purification of PI from *N. alata* stigmas.

- (a) Sephadex G-50 gel filtration chromatography of ammonium sulphate precipitated proteins from stigma extracts. The PI activity eluted late in the profile.
- 5 (b) 20% w/v SDS-polyacrylamide gel (Laemmli, 1970) of fractions across the gel filtration column. The gel was silver stained and molecular weight markers (Pharmacia peptide markers) are in kilodaltons. A protein of about 6kD (arrowed) coelutes with the proteinase inhibitor activity.
- 10 (c) Analysis of PI-containing fractions at different stages of the purification procedure, by SDS-PAGE. Lane 1, crude stigma extract (5µg); Lane 2, stigma proteins precipitated by 80% w/v ammonium sulphate (5µg); Lane 3, PI protein eluted from the chymotrypsin affinity column (1µg).

15 The PI is a 6kD protein and is a major component in unfractionated buffer soluble extracts from stigmas.

Figure 7 is a graphical representation showing hydropathy plots of the PI proteins encoded by the NA-PI-2 clone from *N. alata* and the potato and tomato PI II cDNAs. Values above the line denote hydrophobic regions and values below the line denote hydrophilic regions. The putative signal peptides are shaded. The
20 hydrophobicity profile was generated using the predictive rules of Kyte and Doolittle (1982) and a span of 9 consecutive amino acids.

- (a) Hydropathy profile of the *N. alata* PI protein. The six repeated domains in the predicted precursor protein are labelled I-VI. The hydrophilic regions
25 containing the putative cleavage sites for production of the 6kD PI species are arrowed. The regions corresponding to the peptides that would be produced by cleavage at these sites are marked C for chymotrypsin inhibitor, T for trypsin inhibitor and x for the two flanking peptides.
- (b) Hydropathy profile of the potato PI II protein. (Sanchez-Serrano *et al.*, 1986).
30 The two repeated domains in the PI II protein are labelled I and II. The putative cleavage sites for production of PCI-1 are arrowed (Hass *et al.*, 1982) and the region spanned by PCI-1 is marked.

- 17 -

- (c) Hydropathy profile of the polypeptide encoded by the tomato PI II cDNA (Graham *et al.*, 1985). The two domains are labelled, I and II and the residues which would be potential processing sites are arrowed. These sites are not present in regions predicted to be hydrophilic and consequently a cleavage product is not marked.

Figure 8 shows an immunoblot analysis of the PI protein in stigmas of developing flowers.

- (a) Developing flowers of *N. alata*.
- 10 (b) SDS-PAGE of stigma proteins at the stages of development shown in (a) 5 µg of each extract was loaded. The peptide gel was silver stained and molecular weight markers (LKB Low Molecular weight and Pharmacia peptide markers) are in kilodaltons.
- (c) Immunoblot of a gel identical to (b), probed with anti-PI antiserum.

15

Stigmas from developing flowers contain four proteins of approximately 42kD, 32kD, 18kD and 6kD that bind to the anti-PI antibody. The 42kD and the 18kD components decrease in concentration as the flowers mature, while the 6kD PI protein reaches a maximum concentration just before anthesis. The level of the 32kD component, which runs as a doublet, does not alter significantly during flower development.



20

Figure 9 shows the separation and identification of the 6kD proteinase inhibitor species from *N.alata* stigmas

- 25 A. Separation of the 6kD PIs by reversed phase HPLC chromatography

Four major peaks were obtained with retention times of about 15.5min(peak1), 20.5min(peak2), 22.5min(peak3), 24min(peak4). The peptides in each peak have been identified by a combination of N-terminal analysis and mass spectrometry. See B for description of C1 and T1-T4.

30

B. The five homologous peptides produced from the PI precursor protein: C1, chymotrypsin inhibitor, T1-T4 trypsin inhibitors. The solid bars represent the reactive sites of the inhibitors. The precursor protein is drawn minus the signal sequence.  region of the six repeated domains (amino acids 1-343, Fig.1).  non-repeated sequence (amino acids 344-368, Fig.1). The arrows point to the processing sites in the precursor protein.

C. The amino acid sequence of C1 and T1-T4 predicted from the cDNA clone and confirmed by N-terminal sequencing of the purified peptides. The amino acid at the carboxy-terminus of each peptide was obtained by accurate mass determination using an electro-spray mass spectrometer. The C1 and T1 inhibitors differ by five amino acids (bold). Two of these amino acids are located at the reactive site (underlined) and the other two to three reside at the carboxy-terminus. Peptides T2-T4 have changes in three amino acids (boxed) that are conserved between C1 and T1. Peptides T2 and T3 are identical to each other. Mass spectrometry was used to demonstrate that other forms of C1 and T1-T4 occur due to non-precise trimming at the N- and C-termini. That is, some forms are missing residue 1 or residue 53 and others are missing both residue 1 and 53 (see Table 2).

Figure 10 shows the amino-acid sequence around the processing sites in the precursor PI protein.

The sequence in bold is the amino-terminal sequence obtained from the purified PI protein. The sequence labelled with negative numbers is the flanking sequence predicted from the cDNA clone. The predicted precursor protein contains six repeats of this sequence.

Figure 11 shows the PI precursor produced in a baculovirus expression system and the products obtained after digestion of the affinity purified PI precursor by the endoproteinase Asp-N.

A. The PI precursor produced by the recombinant baculovirus.

Immunoblot containing affinity and HPLC purified PI precursor from *N.alata* stigmas at the green bud stage of development (lane 1) and affinity purified PI precursor produced by the recombinant baculovirus (lane 2). Proteins were fractionated by electrophoresis on a 15% w/v SDS-polyacrylamide gel prior to electrophoretic transfer to nitrocellulose. The blot was incubated with the antibody raised in rabbits to the 6kD PI species from stigmas. The recombinant virus produced an immunoreactive protein of 42kD that is the same size as the PI precursor protein produced by stigmas (arrowed).

B. Cleavage of the PI precursor by endoproteinase Asp-N.

15% SDS-polyacrylamide gel stained with silver containing: 1, PI precursor, produced by baculovirus, incubated without enzyme. 2, enzyme incubated without precursor. 6kD, PI peptides of about 6kD purified from *N.alata* stigmas. 1m, 5m, 30m, reaction products produced after 1, 5 and 30 minutes of incubation. 2h and 24h, reaction products after 2 and 24h of incubation. Peptides of about 6-7kD were detected within one minute of incubation of the precursor with the enzyme. After 24h only peptides of 6-7kD were detected. The bands smaller than 42kD in track 1 are due to truncated forms of the precursor produced by premature termination of translation in the baculovirus expression system.

Figure 12 Preparative chromatography by reversed phase HPLC of the peptides produced from the precursor by Asp-N digestion

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HPLC profile of peptides produced by Asp-N digestion of the PI precursor. The major peaks had a retention time of 19 min (termed Asp-N1) and 21 min (termed Asp-N2). The peptides in these peak fractions (1 & 2) had a slightly slower mobility on SDS-PAGE than the 6kD peptides from stigmas (C, inset). The proteinase inhibitory activity of Asp-N1 and Asp-N2 was tested against trypsin and chymotrypsin.

- 20 -

Figure 13 shows a comparison of the trypsin and chymotrypsin inhibition activity of the PI precursor, PI peptides from stigmas and *in vitro* produced PI peptides from the PI precursor.

PI precursor or PI peptides (0-1.0 μ g) were tested for their ability to inhibit 1.0 μ g of trypsin or chymotrypsin as described in the materials and methods. Inhibitory activity is expressed as the percentage of proteinase activity remaining after the proteinase had been preincubated with the PI with 100% remaining activity taken as the activity of the proteinase preincubated with no PI. Experiments were performed in duplicate and mean values were plotted. Deviation from the mean was 8% or less.

Figure 14 is a graphical representation showing a growth curve for *T. commodus* nymphs reared on control artificial diet, soybean Bowman-Birk inhibitor and *N. alata* PI. The vertical axis represents the mean weight of the crickets in each treatment (+/- standard error) in mg. The horizontal axis represents the week number. The crickets reared on the *N. alata* PI showed a lower mean weight than those reared on both the control diet and the diet containing the soybean inhibitor, throughout the experiment.

SUMMARY OF SEQ ID NOs

5

SEQ ID NO. 1	Nucleotide coding region of <i>N. alata</i> PI precursor
SEQ ID NO. 2	Full length nucleotide sequence of <i>N. alata</i> PI precursor
SEQ ID NO. 3	Amino acid sequence corresponding to SEQ ID NO. 1
10 SEQ ID NO. 4	Residues 1-24 of SEQ ID NO. 2 (peptide 1)
SEQ ID NO. 5	Residues 25-82 of SEQ ID NO. 2 (peptide 2)
SEQ ID NO. 6	Residues 83-140 of SEQ ID NO. 2 (peptide 3)
SEQ ID NO. 7	Residues 141-198 of SEQ ID NO. 2 (peptide 4)
SEQ ID NO. 8	Residues 199-256 of SEQ ID NO. 2 (peptide 5)
15 SEQ ID NO. 9	Residues 257-314 of SEQ ID NO. 2 (peptide 6)
SEQ ID NO. 10	Residues 315-368 of SEQ ID NO. 2 (peptide 7)
SEQ ID NO. 11	N-terminal amino acid sequence of 6kD PI protein
SEQ ID NO. 12	N-terminal amino acid sequence of 6kD PI protein

20

EXAMPLE 1**1. MATERIALS AND METHODS****Plant Material**

- 25 *Nicotiana alata* (Link et Otto) plants of self-incompatibility genotype S_1S_3 , S_3S_3 and S_6S_6 were maintained under standard glasshouse conditions as previously described (Anderson *et al.*, 1989). Organs were collected directly into liquid Nitrogen to avoid induction of a wound response and stored at -70° until required. To study the effect of wounding on gene expression, leaves were wounded by crushing across the mid-
30 vein with a dialysis clip. Leaves were collected 4 and 24 hours after wounding.

Identification and sequencing of a cDNA clone encoding PI

Polyadenylated RNA was prepared from stigmas and styles, isolated from mature flowers of *N. alata* (genotype S_3S_3), and used to construct a cDNA library in Lambda gt10 (Anderson *et al.*, 1989). Single stranded ^{32}P -labelled cDNA was prepared from mRNA from stigmas and styles of *N. alata* (genotype S_3S_3 and S_6S_6) and used to screen the library for highly expressed clones which were not S-genotype specific (Anderson *et al.*, 1989). Plaques which hybridised strongly to cDNA probes from both S-genotypes were selected and assembled into groups on the basis of cross-hybridisation. The longest clone of each group was subcloned into M13mp18 and pGEM 3zf+, and sequenced using an Applied Biosystems Model 373A automated sequencer. Both dye primer and dye terminator cycle sequencing chemistries were performed according to standard Applied Biosystems protocols. Consensus sequences were generated using SeqEdTM sequence editing software (Applied Biosystems). The GenBank database was searched for sequences homologous to these clones. Because of the high degree of sequence similarity between the six domains of the *N. alata* PI clone, sequencing primers were made to non-repeated 3' sequences (nucleotides 1117-1137, 1188-1203 and 1247-1267), and to a 5' sequence before the start of the repetitive regions (nucleotides 74-98). In addition, the pNA-PI-2 insert was restricted with endonuclease *Hae*II, which cut at nucleotides 622 and 970 to produce three fragments. The fragments were subcloned into pGEM7zf+ and sequenced in both directions, using the M13 forward and reverse primers. The repetitive nature of the pNA-PI-2 insert rendered it unstable in both phagemid and plasmid vectors when cultures were grown longer than 6 hours.

25

RNA Gel Blot Analysis

Total RNA was isolated and separated on a 1.2% w/v agarose/formaldehyde gel as previously described (Anderson *et al.*, 1989). The RNA was transferred to Hybond-N (Amersham) and probed with the insert from pNA-PI-2 labelled with ^{32}P using random hexanucleotides (1×10^8 cpm μg^{-1} ; 1×10^7 cpm ml^{-1}) (Feinberg and Vogelstein, 1983). Prehybridisation and hybridisation, at 68 °C, were as described by Anderson *et al.* (1989). The filters were washed in 2 x SSC, 0.1% w/v SDS or 0.2

30

x SSC, 1% w/v SDS at 68 °C.

***In situ* hybridisation**

In situ hybridisation was performed as described by Cornish *et al*, 1987. The probe
5 was prepared by labelling the insert from pNA-PI-2 (100ng) to a specific activity of
 10^8 cpm μg^{-1} by random hexanucleotide priming (Feinberg and Vogelstein, 1983).
The labelled probe was precipitated, and resuspended in hybridisation buffer (50 μl),
and 5 μl was applied to the sections. The sections were covered with coverslips, and
incubated overnight at 40 °C in a closed box containing 50% v/v formamide. After
10 incubation, sections were washed sequentially in 4 x SSC at room temperature, 2 x
SSC at room temperature, and 1 x SSC at 40 °C for 40 min. The slides were dried
and exposed directly to X-ray film (Cronex MRF 32, Dupont) at room temperature,
overnight. Hybridised sections were counterstained with 0.025% w/v toluidine blue
in H₂O, and mounted in Eukitt (Carl Zeiss, Freilburg, FRG). Autoradiographs were
15 transposed over sections to give the composites shown.

DNA Gel Blot Analysis

Genomic DNA was isolated from young leaves of *N. alata* by the procedure of
Bernatzky and Tanksley (1986). DNA (10 μg) was digested to completion with the
20 restriction endonucleases *Eco*RI or *Hind*III, separated by electrophoresis on a 0.9%
w/v agarose gel, and transferred to Hybond-N (Amersham) by wet blotting in 20 x
SSC. Filters were probed and washed as described for RNA blot analysis.

Preparation of protein extracts

25 Soluble proteins were extracted from plant material by freezing the tissue in liquid
N₂, and grinding to a fine powder in a mortar and pestle. The powdered tissue was
extracted in a buffer consisting of 100mM Tris-HCl, pH 8.5, 10mM EDTA, 2mM
CaCl₂, 14 μM β -mercaptoethanol. Insoluble material was removed by centrifugation
at 10,000g for 15 min. Protein concentrations were estimated by the method of
30 Bradford (1976) with Bovine Serum Albumin (BSA) as a standard.

Proteinase inhibition assays

Protein extracts and purified protein were assayed for inhibitory activity against trypsin and chymotrypsin as described by Rickauer *et al* (1989). Inhibitory activity
5 was measured against 1 µg of trypsin (TPCK-treated; Sigma) or 3 µg of chymotrypsin (TLCK-treated; Sigma). The rate of hydrolysis of synthetic substrates N- α -P-tosyl-L-arginine methyl ester (TAME) and N-benzoyl-L-tyrosine ethyl ester (BTEE) by trypsin and chymotrypsin, respectively, were taken as the uninhibited activity of the
10 enzymes. Inhibitory activity of the extract was expressed as the percentage of control protease activity remaining after the protease had been pre-incubated with the extract. The PI peptides from stigma, PI precursor and Asp-N processed peptides were assayed for inhibitory activity as described by Christeller *et al* (1989).

Purification of the *N. alata* PI protein

15 Stigmas (1000; 10g) were ground to a fine powder in liquid N₂ and extracted in buffer (100mM Tris-HCl, pH8.5, 10mM EDTA, 2mM CaCl₂, 14µM β -mercaptoethanol, 4ml/g tissue). To concentrate the extract prior to the first purification step, gel filtration, the inhibitory activity was precipitated with 80% w/v ammonium sulphate, the concentration required to precipitate all the proteinase
20 inhibitory activity.

The ammonium sulphate pellet was resuspended in 5ml of 0.15M KCl, 10mM Tris-HCl, pH 8.1, and loaded onto a Sephadex G-50 column (2cm x 100cm) equilibrated with the same buffer. The fractions (10ml) eluted from this column and containing
25 proteinase inhibitory activity were pooled and applied to an affinity column of Chymotrypsin-Sepharose CL4B [100mg TLCK-treated α -chymotrypsin (Sigma) cross-linked to 15ml Sepharose CL4B (Pharmacia) by manufacturers instructions]. The column was washed with 10 volumes of 0.15M KCl/10mM Tris-HCl, pH 8.1, prior to elution of bound proteins with 7m urea, pH 3 (5 ml fractions). The eluate was
30 neutralised immediately with 200 µl 1M Tris-HCl pH 8, and dialyzed extensively against deionised H₂O.

Amino acid sequence analysis

Purified PI protein was chromatographed on a reverse phase HPLC microbore column prior to automated Edman degradation on a gas phase sequencer (Mau *et al.*, 1986). Phenylthiohydantoin (PTH) amino acids were analysed by HPLC as
5 described by Grego *et al.* (1985).

Production of a polyclonal antiserum to the *N. alata* PI

The purified proteinase inhibitor (Figure 6c, lane 3) was conjugated to a carrier protein, keyhole limpet haemocyanin (KLH) (Sigma), using glutaraldehyde, as
10 follows. 1mg of PI protein was dissolved in 1.5ml H₂O, and mixed with 0.3 mg KLH in 0.5 ml of 0.4M phosphate buffer, pH7.5. 1ml of 20mM glutaraldehyde was added dropwise over 5 min, with stirring at room temperature. The mixture was stirred for 30 min at room temperature, 0.25ml of glycine was added, and the mixture was stirred for a further 30 min. The conjugated protein was then dialyzed extensively
15 against normal saline (0.8% w/v NaCl). The equivalent of 100µg of PI protein was used for each injection. Freund's complete adjuvant was used for the first injection, and incomplete adjuvant for two subsequent booster injections. The IgG fraction of the antiserum was separated on Protein A Sepharose (Pharmacia) according to manufacturer's instructions.

20

Protein Gel Blot Analysis

Protein extracts were electrophoresed in 15% w/v SDS-polyacrylamide gels (Laemmli, 1970) and transferred to nitrocellulose in 25mM Tris-HCl, 192mM glycine, 20% v/v methanol, using a BioRad Trans-Blot^RSemi-dry electrophoretic transfer cell
25 (12V, 20 min). Loading and protein transfer were checked by staining the proteins on the membranes with Ponceau S (Harlow and Lane, 1988). Membranes were blocked in 3% w/v bovine serum albumin for 1h, and incubated with the anti-PI antibody (2µg/ml in 1% w/v BSA, Tris Buffered Saline) overnight at room temperature. Bound antibody was detected using biotinylated donkey anti-rabbit IgG
30 (1/500 dilution, Amersham) and the Amersham Biotin-Streptavidin system according to procedures recommended by the manufacturer.

- 26 -

Proteolysis of the PI precursor by endoproteinase Asp-N

Affinity-purified PI precursor (1.25mg) was incubated at 37°C with endoproteinase Asp-N (2µg) in 100mM NH₄HCO₃, pH 8.5 in a total volume of 1ml for 48h. Reaction products were separated by reversed-phase HPLC using an analytical
5 Brownlee RP-300 Aquapore column (C8, 7µm, 4.6x100mm). The column was equilibrated in 0.1% v/v TFA and peptides were eluted with the following program: 0-25%B (60% v/v acetonitrile in 0.089% v/v TFA) applied over 5min, followed by a gradient of 25-42%B over the next 40min, and ending with a gradient of 42-100%B over 5 minutes. The flow rate was 1.0ml/min and peptides were detected by
10 absorbance at 215nm. Each peak was collected manually and freeze dried. Concentration was estimated by response obtained with each peak on the UV detector at 215nm.

2. CLONING OF PI PRECURSOR GENE

15

Isolation and characterisation of the PI cDNA clone

A cDNA library, prepared from mRNA isolated from the stigmas and styles of mature flowers of *N. alata*, was screened for clones of highly expressed genes which were not associated with self-incompatibility genotype. Clones encoding a protein
20 with some sequence identity to the type II proteinase inhibitors from potato and tomato (Thornburg *et al.*, 1987; Graham *et al.*, 1985) were selected. The largest clone, NA-PI-2, is 1360 base pairs long with an open reading frame of 1191 nucleotides. The nucleic acid sequence (SEQ ID NO. 2) and the predicted amino acid sequence (SEQ ID NO. 3) of the *N. alata* clone, NA-PI-2 is shown in Figure 1.
25 There are no potential N-glycosylation sites.

Surprisingly, the *N. alata* cDNA clone encodes a protein with six repeated domains that have high, but not perfect, sequence identity (Figure 1). Each of these domains contains a potential reactive site which is highlighted in Figure 1. The residues at
30 the putative reactive sites of the *N. alata* PI are consistent with the inhibitor having two sites which would specifically inhibit chymotrypsin (Leu5-Asn6, Leu63-Asn64) and four sites specific for trypsin (Arg121-Asn122, Arg179-Asn180, Arg237-Asn238

- 27 -

and Arg295-Asn296).

To ensure that the repeat structure of NA-PI-2 was not due to a cloning artifact, three additional cDNA clones were sequenced, and found to be identical to NA-PI-2.

5

Table 1 is a comparison of the percentage amino acid identity of the six domains of the PI precursor.

Temporal and spatial expression of the PI mRNA

- 10 Total RNA, isolated from various tissues of *N. alata*, was probed with the PI cDNA clone in the RNA gel blot analyses shown in Figure 2. Two hybridising messages of 1.0 and 1.4kb were present in total RNA isolated from styles (including stigmas). Only the larger message, which was predominant in this tissue, is of sufficient size to encode the cDNA clone NA-PI-2 (1.4kb). The smaller message is not detected
- 15 with the cDNA probe at higher stringency. An homologous message of approximately 1.4kb was also present in RNA isolated from the styles of *N. tabacum* and *N. sylvestris* (Figure 2).

- In the other floral organs (except pollen), both messages were detectable at low
- 20 levels, however, the smaller RNA species appeared more abundant. There was no hybridisation to pollen RNA. No hybridising species were evident in leaf RNA, but two species, 1.0 and 1.4kb were detected 24 hours after mechanical wounding. The smaller message (1.0kb) was more abundant in this case.

- 25 *In situ* hybridisation of radiolabelled *N. alata* PI cDNA to longitudinal sections of styles from immature (1cm long) buds is shown in Figure 3. RNA homologous to the cDNA clone bound strongly to cells of the stigma and weakly to vascular bundles. No hybridisation was detected in the cortical tissue, transmitting tract tissue, or epidermis of the style. The same pattern of hybridisation was observed in
- 30 mature receptive flowers. Control sections treated with ribonuclease A prior to hybridisation were not labelled.

Genomic DNA blot analysis

The cDNA clone NA-PI-2, was used as a probe on the DNA gel blot shown in Figure 4 which contained genomic DNA, digested with either *EcoRI* or *HindIII*. *EcoRI* produced two hybridising fragments (11kb and 7.8kb) and *HindIII* produced
5 three large hybridising fragments (16.6, 13.5 and 10.5 kb).

Distribution of PI activity in various tissues of *N. alata*

The inhibition of trypsin and chymotrypsin by crude extracts of various organs of *N. alata* is shown in Figure 5. Stigma extract was the most effective inhibitor of both
10 trypsin and chymotrypsin. The stigma extracts had up to eight times more inhibitory activity than sepal extracts, and more than 20 times more activity than extracts from styles, petals, leaves and wounded leaves.

Purification of PI from *N. alata* stigmas

15 Stigmas of *N. alata* were extracted in buffer and the inhibitory activity was concentrated by precipitation with 80% w/v ammonium sulphate. The precipitate was redissolved and fractionated by gel filtration on Sephadex G-50. Most of the protein in the extract eluted early in the profile illustrated in Figures 6a and 6b, relative to the proteinase inhibitor. Fractions with proteinase inhibitor activity were
20 pooled and applied to an affinity column of chymotrypsin-Sepharose. The PI activity co-eluted with a protein of about 6kD, which appeared to migrate as a single band on the 20% SDS-polyacrylamide gel shown in Figure 6c. The purity of the PI at various stages of purification was assessed by SDS-PAGE (Figure 6c). The purified inhibitor represented approximately 50% of the inhibitory activity present in the
25 crude extract.

Amino acid sequence of the N-terminus of the 6kD PI protein

The N-terminal amino acid sequence DRICTNCCAG(T/K)KG (SEQ ID NO. 11; SEQ ID NO. 12, respectively) was obtained from the purified PI protein. This
30 sequence of amino acids corresponds to six regions in the deduced sequence of the cDNA clone, starting at positions 25, 83, 141, 199, 257 and 315 in Figure 1. At position 11 of the N-terminal sequence, both threonine and lysine were detected.

- 29 -

This is consistent with the purified inhibitor comprising a mixture of six peptides beginning with the sequences underlined in Figure 1, as the first two peptides contain threonine at this position, while the other four peptides have lysine at this position. The position of these peptides relative to the six repeated domains in the predicted precursor protein is illustrated in Figure 7. Five of the six predicted 6kD peptides, contain a reactive site for either chymotrypsin or trypsin (Figure 1 and 7). The sixth potential peptide is four amino-acids shorter than the other five peptides (fifty eight amino-acids) and may not be active, as it does not contain an inhibitory site. The peptide from the N-terminus (x in Figure 7) has a potential chymotrypsin reactive site but is much shorter (24 amino acids).

Distribution of the PI protein in *N. alata*

A polyclonal antiserum was raised to the purified PI protein conjugated to keyhole limpet haemocyanin. The antibody reacted strongly with the purified 6kD PI protein in immunoblot analyses and bound only to a 6kD and a 32kD protein, which appears as a doublet, in total stigma and style extracts from mature flowers. Figure 8 is an immunoblot containing protein extracts of stigmas from flowers at different stages of development (1cm long buds to mature flowers) probed with the anti-PI antiserum. Larger cross reacting proteins of approximately 18kD, and 42kD were detected in buds from 1cm to 5cm in length in addition to the 6kD and the 32kD protein. The 18kD and 42kD proteins decreased in concentration with maturity, while the 6kD protein reached a peak concentration just before anthesis. The concentration of the 32kD protein remained relatively constant during flower maturation.

25

- 30 -

TABLE 1

<i>N. alata PI</i>							
		1	2	3	4	5	6
<i>N. alata</i>	1		100	88	88	90	79
	2			88	88	90	79
	3				97	95	86
	4					98	90
	5						90
	6						

EXAMPLE 2

PURIFICATION AND IDENTIFICATION OF PI MONOMERS

1. MATERIALS AND METHODS

Separation of the 6kD PI species by reversed phase chromatography

Stigmas (21,000) were ground and extracted as described for purification of the PI protein. After gel filtration on a Sephadex G-50 gel filtration column (5cmx800cm, 3000 stigmas per separation) the peptides were lyophilized and applied to a Brownlee RP-300 C8 Reversed-phase column, 10x250mm, on a Beckman HPLC system Gold, and eluted with 0.1% v/v Trifluoroacetic acid (TFA) and an acetonitrile gradient (0-10% over 5mins, 10-25% over 40 mins and 25-60% over 10 mins), at 5ml/min. Peak fractions, designated fraction 1, 2, 3 and 4 were collected and freeze dried.

Electrospray mass spectrometry

On line mass spectrometric analysis of HPLC eluates was performed by application of 20 pmoles of each PI preparation (fraction 1, 2, 3 & 4) in 2µl of water onto a Brownlee RP-300 C8 reversed-phase column (150x0.20mm internal diameter fused-silica capillary column) on a modified Hewlett- Packard model HP1090L liquid chromatograph and elution with a linear gradient of acetonitrile (0.05% v/v TFA to 0.045% v/v TFA/60% v/v acetonitrile in 30 min.) at a flow rate of 1µl/min and a column temperature of 25°C. The eluant was monitored at 215nm using a Spectral Physics forward optics scanning detector with a 6-mm pathlength U-shaped axial beam capillary flow cell (LC Packings, Netherlands). Mass spectra were acquired on a Finnigan-Mat triple quadrupole mass spectrometer (model TSQ-700, San Jose, CA) equipped with an electrospray ionisation (ESI) source (Analytica, Branford, CT). The electrospray needle was operated in positive ion mode at a voltage differential of -4 kV. The sheath liquid was 2-methoxyethanol delivered at 1µl/min via a syringe drive (Harvard Apparatus, South Natick, MA). The nitrogen drying gas conditions were as follows: heater temperature, 275°C; pressure, 15 psi; flow rate, ~ 15stdL/min. The nitrogen sheath gas was supplied at 33 psi. Gaseous nitrogen was

obtained from a boiling liquid nitrogen source. Peptides were introduced into the ESI source at 1.0 $\mu\text{l}/\text{min}$ by on-line capillary RP-HPLC as described above. Spectra were acquired scanning from m/z 400 to 2000 at a rate of 3sec. Data collection and reduction were performed on a Dec5100 computer using Finnigan BIOMASSTM software.

2. RESULTS

Separation and identification of the individual 6kD PI species from *N.alata* stigmas

The five of the six peptides of about 6kD that were predicted to be present in the purified 6kD PI preparation have been separated from each other by reversed-phase HPLC chromatography. Four peaks were obtained (Fig. 9a) and the peptides within each peak were identified by electrospray mass spectrometry (Table 2). The peptides have been designated C1, T1, T2, T3 and T4 according to their position in the PI precursor and the presence of a chymotrypsin or trypsin reactive site (Fig 9b). The first HPLC peak (Fig 9a) corresponds to the chymotrypsin inhibitor C1, the second peak is composed of a mixture of T2 and T3 (identical to each other) and T4 that differs from T2 and T3 by one amino-acid at position 32. The third peak contains the peptide T1 and the fourth peak is composed of a mixture of T1, T2/T3 and T4 (Table 2).

The site of processing has not been precisely determined, but is likely to be located between the aspartate (N) and asparagine (D) residues in the sequence outlined in Figure 10. Proteases with specific requirements for asparagine residues have been isolated from vacuoles from immature soybean seeds and pumpkin cotyledons (Scott et al., 1992, Hara-Nishimura et al., 1991). This is consistent with the immunogold localization of the PI in the vacuoles of the papillae and the underlying secretory cells in the stigma of *N.alata* (Atkinson, 1992). In the case of the *N.alata* PI, processing analogous to that of peptide hormones is also possible because each of the possible 6kD peptides are flanked by dibasic residues (Lys-Lys, position -2 & -3 in Figure 10). However, a system like this has not been described in plants, and it is more likely that the dibasic residues contribute to the predicted hydrophilic loops

that present the processing site on the surface of the molecule.

The data from the mass spectrometric analysis shows that once the initial cleavage has occurred the new carboxy terminus is trimmed back (Figure 10). The EEKKN sequence (SEQ ID NO. 14) is removed completely but the trimming is not precise, sometimes an additional amino acid is removed. Steric hindrance probably prevents further trimming. Occasionally the aspartate is also removed from the N-terminus.

EXAMPLE 3

Production of PI precursor in insect cell (Sf9) culture using a recombinant baculovirus vector.

cDNA encoding the PI precursor (Figure 1) was inserted into the Eco R1 site of the plasmid vector pVL 1392, which is the same as pVL941 (Lucknow and Summers, 1989) except that a multiple cloning site was inserted at the BamH1 site. The plasmid designated pRH11, contains the PI cDNA in the correct orientation with respect to the direction of transcription directed by the polyhedrin promoter. Recombinant baculovirus was obtained by co-transfection of *Spodoptera frugiperda* cells with baculovirus DNA and pRH11. The recombinant viruses, produced by homologous recombination, were plaque purified and amplified prior to infection of insect cells for protein production. All procedures for production of recombinant baculovirus, titration of the virus and maintenance and infection of the Sf9 cells were obtained from King and Posse (1992). For production of the PI precursor, monolayers of Sf9 cells in large flasks (175cm²) were infected at the time of confluence with an inoculum of high-titre recombinant virus at a multiplicity of infection of 5-10 pfu/cell. Culture fluid was collected after 4 days of infection, clarified by centrifugation and the PI precursor was purified by application to a Chymotrypsin-Sepharose affinity column as described for the 6kD PI species from stigmas. PI precursor eluted from the column in 7M urea, pH3 was neutralized immediately with 1M Tris-HCl buffer pH8, dialysed extensively against Milli-Q water, concentrated 20-50 fold by ultrafiltration using a Diaflow YM10 filter and stored frozen at -20°C.

- 34 -

The cDNA clone encoding the PI precursor was engineered into a baculovirus vector for the production of the precursor from infected insect cells. The insect cells produced a 42kD protein that cross reacted with the antibodies raised to the 6kD PI peptides from stigma and bound to the chymotrypsin affinity column. This 42kD protein was identical in size to the 42kD precursor produced in the immature stigmas of *N.alata* (Fig.11) and had the N-terminal sequence LysAlaCysThrLeuAsn (SEQ ID NO. 13) demonstrating that the signal sequence had been processed correctly by the insect cells (Fig.1). Based on these results, the 42kD protein produced in the baculovirus expression system will now be referred to as the PI precursor. The 42kD PI precursor had inhibitory activity against chymotrypsin but no inhibitory activity against trypsin (Fig.13). Processing of the PI precursor by the endoproteinase AspN led to the production of stable peptides of about 6kD that were partially purified by reversed phase HPLC (Fig.12). These peptides have equivalent inhibitory activity against trypsin and chymotrypsin as the 6kD peptides isolated from stigma, indicating that processing of the precursor is required to activate the trypsin inhibitory activity but not all the chymotrypsin activity. Since AspN cleaves specifically adjacent to Aspartate residues (between Asn-1 and Asp1 in Figure 10) and has no trimming activity, the peptides produced *in vitro* will be similar to those produced in stigmas except for the presence of the sequence EEKKN (SEQ ID NO. 14) at the C-terminus. This provides further evidence that precise processing of the N-and C-termini is not required to obtain an active 6kD PI peptide. Asp-N1 is more efficient at inhibiting chymotrypsin than trypsin and is thus likely to be predominantly a C1 analogue (Fig.9b). Asp-N2 is a more efficient trypsin inhibitor and probably contains the T1-T4 analogues.

EXAMPLE 4

Effect of PIs on protease activity in unfractionated gut extracts from various insects

Activity of PIs on gut proteases was measured using the procedure of Christeller et al., (1992) as follows. An aliquot of 1 μ M of inhibitor (0-10 μ l, at least 5-fold excess over proteases present in the gut) was mixed with 150 μ l of 10mM CAPS buffer, pH 10, and preincubated with each insect gut extract (0-15 μ l), for 20 min at 30°C. The reaction was started by the addition of 50 μ l of 14 C -labelled casein substrate (400 μ g protein, specific activity 25,000-75,000 dpm mg $^{-1}$) and continued for 30 min at 30°C until 50 μ l of cold 30% (w/v) TCA was added to terminate the reaction. After incubation on ice for 30 min, undigested protein was pelleted by centrifugation at 20°C for 5 min at 10,000g. The supernatant was removed, mixed with scintillation fluid and the radioactivity measured. Assays were performed at pH 10 except for *L.sericata* and *C.rufifacies* when 10mM Tris-HCl, pH 8.0 was used.

Table 3 shows the inhibitory activity of the pooled 6kD PI peptides (C1, T1, T2/T3, T4), the mixture of trypsin inhibitors T2/T3 and T4, and the chymotrypsin inhibitor C1 against the proteases in the gut of various members of the Lepidoptera, Coleoptera, Orthoptera and Diptera. In most cases, the pooled peptides and the trypsin inhibitors had an equivalent effect against the gut proteases with the degree of inhibition ranging from 37-79% depending on the insect tested. The inhibitors had negligible effect on the gut proteases of the potato tuber moth, *P.opercullela*. The chymotrypsin inhibitor C1 also affected the activity of the proteases but was less effective than the trypsin inhibitors in five cases (*W.cervinata*, *L.sericata*, *C.zealandica*, *P.octa*, sugar cane grub).

The experimental details are described in the legend to Figure 14. The *N. alata* PI was more effective than Soybean Bowman-Birk inhibitor in reducing cricket weight. It has shown that there is a good correlation between the ability of a proteinase inhibitor to inhibit the enzymes of the insect midgut and its effectiveness in retarding the growth of insects in insect feeding trials (Christeller et al., 1992). Figure 14

- 36 -

shows that the pooled PIs that inhibited the gut proteases of the black field cricket (*T.commodus*) by 70% in the *in vitro* assay retarded the growth of the crickets by 30% in a feeding trial conducted over a 10 week period. The correlation between *in vitro* assays and feeding trials has been confirmed recently by Johnston and colleagues (1993) working on growth and development of *Helicoverpa armigera*.

- 37 -

TABLE 2

HPLC peak	retention time (min)	molecular weight	assigned peptide*
1	15.5	5731.5	C1
		5644.4	C1 minus Ser ₅₃
		5616.4	C1 minus Asp ₁ & Ser ₅₃
		55.29.3	C1 minus Asp ₁
2	20.5	5700.5	T2/T3
		5728.5	T4
		5585.4	T2/T3 minus Asp ₁
		5613.5	T4 minus Asp ₁
3	22.5	5725.5	T1
		5610.5	T1 minus Asp ₁
4	24	5654.4	T1 minus Ala ₅₃
		5641.4	T4 minus Ser ₅₃
		5613.4	T2/T3 minus Ser ₅₃
		5539.4	T1 minus Asp ₁ & Ala ₅₃
		5498.4	T2/T3 minus Asp ₁ & Ser ₅₃
		5526.4	T4 minus Asp ₁ & Ser ₅₃

* See Figure 9 for designation of C1 and T1-T4.

TABLE 3
Effect of *Nicotiana glauca* proteinase inhibitors and Potato
inhibitor II on casein hydrolysis by crude gut extracts

	casein hydrolysis (% control)		
Insect	NaPI	C1	T2/T3, T4
<i>H. armigera</i>	33.2	32.7	30.3
<i>H. punctigera</i>	26.6	29.3	28.5
<i>T. commodus</i>	28.4	35.0	33.1
<i>A. infusa</i>	37.5	40.2	43.3
sugar cane grub	25.8	43.9	25.1
<i>W. cervinata</i>	22.9	82.9	20.4
<i>E. postvitiana</i>	39.7	45.4	41.2
<i>S. litura</i>	28.1	33.6	24.8
<i>P. opercullela</i>	95.8	100	98.5
<i>C. rufifacies</i>	29.1	37.8	28.9
<i>L. serricata</i>	59.2	100	63.0
<i>C. zealandica</i>	31.7	54.7	32.0
<i>P. octo</i>	57.1	67.2	57.4
<i>C. obliquana</i>	51.1	49.1	45.5
<i>A. tasmaniae</i>	28.3	34.2	39.5

Legend to Table 3

NaPI = *N. alata* proteinase inhibitors pooled

C1 = *N. alata* chymotrypsin inhibitor (peak 1 from HPLC)

T2/T3, T4 = *N. alata* trypsin inhibitors (peak 2 from HPLC)

Heliothis armigera, *Helicoverpa armigera*, Tobacco budworm, Lepidoptera

Heliothis punctigera, *Helicoverpa punctigera* Native budworm, Lepidoptera

Teleogryllus commodus Black field cricket, Orthoptera

Agrotis infusa Common cutworm, adults known as the Bogong moth, Lepidoptera

Wiseana cervinata Porina, native to New Zealand, Lepidoptera

Lucilla sericata Green blow fly, Diptera, assayed at pH8

Chrysomya rufifacies Hairy maggot blow fly, Diptera, assayed at pH8

Aphodius tasmaniae Tasmanian grass grub = Black-headed pasture cockchafer, Coleoptera

Costelytra zealandica New Zealand grass grub, Coleoptera

Spodoptera litura Tropical armyworm, Lepidoptera

Phthorimaea operculella Potato tuber moth, Lepidoptera

Epiphyas postvittana Lightbrown apple moth (leafroller), Lepidoptera

Planotortrix octo Greenheaded leafroller, Lepidoptera

Ctenopseustis obliquana Brownheaded leafroller, Lepidoptera

Sugar cane grub

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and compounds referred to or indicated in this specification, individually or collectively, and any and all combinations of any two or more of said steps or features.

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- 43 -

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: (other than US) THE UNIVERSITY OF MELBOURNE
(US only) ANDERSON, M A; ATKINSON, A H;
HEATH, R L; and CLARKE, A E.
- (ii) TITLE OF INVENTION: A PROTEINASE INHIBITOR, PRECURSOR
THEREOF AND GENETIC SEQUENCES ENCODING
SAME
- (iii) NUMBER OF SEQUENCES:
- (iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: DAVIES COLLISON CAVE
(B) STREET: 1 LITTLE COLLINS STREET
(C) CITY: MELBOURNE
(D) STATE: VICTORIA
(E) COUNTRY: AUSTRALIA
(F) ZIP: 3000
- (v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER: AU PCT International
(B) FILING DATE: 16-DEC-1993
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(A) APPLICATION NUMBER: PL6399 - Australia
(B) FILING DATE: 16-DEC-1992
- (viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: SLATTERY, JOHN M
(C) REFERENCE/DOCKET NUMBER: EJH/JMS/EK
- (ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: (613) 254 2777
(B) TELEFAX: (613) 254 2770

- 44 -

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1104 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

BNSDOCID: <WO 9413810A1>

- 46 -

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135 140 145 150	
AAA AAG GGT TGT AAG TAC TTT AGT GAT GAT GGA ACT TTT GTT TGT GAA	594
Lys Lys Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr Phe Val Cys Glu	
155 160 165	
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Gly Glu Ser Asp Pro Lys Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp	
170 175 180	
GGA AGA ATT GCC TAT GGG ATT TGC CCA CTT TCA GAA GAA AAG AAG AAT	690
Gly Arg Ile Ala Tyr Gly Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn	
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Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr	
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Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Lys	
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AAT CCA AAG GCT TGT CCT CGG AAT TGT GAT GGA AGA ATT GCC TAT GGG	834
Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly	
235 240 245	
ATT TGC CCA CTT TCA GAA GAA AAG AAG AAT GAT CGG ATA TGC ACA AAC	882
Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn	
250 255 260	

- 47 -

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- 48 -

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 368 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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 Ala Gly Thr Lys Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr Phe Val
 35 40 45
 Cys Glu Gly Glu Ser Asp Pro Arg Asn Pro Lys Ala Cys Thr Leu Asn
 50 55 60
 Cys Asp Pro Arg Ile Ala Tyr Gly Val Cys Pro Arg Ser Glu Glu Lys
 65 70 75 80
 Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Thr Lys Gly Cys
 85 90 95
 Lys Tyr Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp
 100 105 110
 Pro Arg Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Pro Arg Ile Ala
 115 120 125
 Tyr Gly Ile Cys Pro Leu Ala Glu Glu Lys Lys Asn Asp Arg Ile Cys
 130 135 140
 Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr Phe Ser Asp Asp
 145 150 155 160
 Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Lys Asn Pro Lys Ala
 165 170 175
 Cys Pro Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly Ile Cys Pro Leu
 180 185 190

- 49 -

Ser Glu Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly
 195 200 205

Lys Lys Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr Phe Val Cys Glu
 210 215 220

Gly Glu Ser Asp Pro Lys Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp
 225 230 235 240

Gly Arg Ile Ala Tyr Gly Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn
 245 250 255

Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr
 260 265 270

Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Arg
 275 280 285

Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly
 290 295 300

Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn
 305 310 315 320

Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr
 325 330 335

Phe Ile Cys Glu Gly Glu Ser Glu Tyr Ala Ser Lys Val Asp Glu Tyr
 340 345 350

Val Gly Glu Val Glu Asn Asp Leu Gln Lys Ser Lys Val Ala Val Ser
 355 360 365

- 50 -

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 24 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Lys Ala Cys Thr Leu Asn Cys Asp Pro Arg Ile Ala Tyr Gly Val Cys
1 5 10 15
Pro Arg Ser Glu Glu Lys Lys Asn
20

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 58 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Thr Lys Gly Cys Lys Tyr
1 5 10 15
Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Arg
20 25 30
Asn Pro Lys Ala Cys Thr Leu Asn Cys Asp Pro Arg Ile Ala Tyr Gly
35 40 45
Val Cys Pro Arg Ser Glu Glu Lys Lys Asn
50 55

- 51 -

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 58 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Thr Lys Gly Cys Lys Tyr
 1 5 10 15

Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Arg
 20 25 30

Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Pro Arg Ile Ala Tyr Gly
 35 40 45

Ile Cys Pro Leu Ala Glu Glu Lys Lys Asn
 50 55

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 58 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr
 1 5 10 15

Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Lys
 20 25 30

Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly
 35 40 45

Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn
 50 55

- 52 -

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 58 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr
 1 5 10 15

Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Lys
 20 25 30

Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly
 35 40 45

Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn
 50 55

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 58 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr
 1 5 10 15

Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Arg
 20 25 30

Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly
 35 40 45

Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn
 50 55

- 53 -

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 54 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr
1 5 10 15
Phe Ser Asp Asp Gly Thr Phe Ile Cys Glu Gly Glu Ser Glu Tyr Ala
20 25 30
Ser Lys Val Asp Glu Tyr Val Gly Glu Val Glu Asn Asp Leu Gln Lys
35 40 45
Ser Lys Val Ala Val Ser
50

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 13 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Thr Lys Gly
1 5 10

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 13 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly
1 5 10

- 54 -

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 6 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Lys Ala Cys Thr Leu Asn
1 5

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 5 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Glu Glu Lys Lys Asn
1 5

CLAIMS:

1. A nucleic acid isolate comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a type II serine proteinase inhibitor (PI) precursor from a plant wherein said precursor comprises at least three PI monomers and wherein at least one of said monomers has a chymotrypsin specific site and at least one other of said monomers has a trypsin specific site.
2. A nucleic acid isolate according to claim 1 wherein said PI precursor comprises at least four monomers.
3. A nucleic acid isolate according to claim 1 wherein the PI precursor comprises at least five monomers.
4. A nucleic acid isolate according to claim 1 wherein the PI precursor comprises at least six monomers.
5. A nucleic acid isolate according to claim 1 wherein said isolate comprises a sequence of nucleotides as set forth in SEQ ID NO. 1 or having at least 55% nucleotide similarity to all or part thereof.
6. A nucleic acid isolate according to claim 1 or 5 wherein said nucleic acid isolate is capable of hybridising under low stringency conditions to a complementary sequence to SEQ ID NO. 1.
7. A nucleic acid isolate comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a single type II serine PI having either a chymotrypsin specific site or a trypsin specific site and wherein said PI is a monomer of a precursor PI having at least three monomers of which at least one of said monomers has a chymotrypsin site and the other of said monomers has a trypsin site.

- 56 -

8. A nucleic acid isolate according to claim 7 comprising a sequence of nucleotides which is at least 55% similar to all or part of SEQ ID NO. 1.
9. A nucleic acid isolate according to claim 7 which is capable of hybridising under low stringency conditions to a complementary nucleotide sequence to SEQ ID No. 1.
10. A nucleic acid isolate according to claim 7 or 8 or 9 comprising a nucleotide sequence which encodes a peptide selected from (SEQ ID NO. 5); (SEQ ID NO. 6); (SEQ ID NO. 7); (SEQ ID NO. 8); (SEQ ID NO. 9); (SEQ ID NO. 10).
11. A nucleic acid isolate according to claim 7 or 8 or 9 comprising a nucleotide sequence which encodes a peptide defined by SEQ ID NO. 4.
12. A recombinant type II serine PI precursor from a plant wherein said precursor comprises at least three PI monomers and wherein at least one of said monomers has a chymotrypsin site and at least one other of said monomers has a trypsin specific site.
13. A recombinant PI precursor according to claim 12 wherein said PI precursor comprises at least four monomers.
14. A recombinant PI precursor according to claim 12 wherein said PI precursor comprises at least five monomers.
15. A recombinant PI precursor according to claim 12 wherein said PI precursor comprises at least six monomers.
16. A recombinant PI precursor according to claim 12 wherein said PI precursor comprises a sequence of amino acids as set forth in SEQ ID NO. 3 or having at least 55% similarity to all or part thereof.

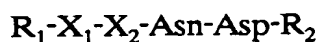
- 57 -

17. A monomer of the recombinant PI according to claim 12.

18. A monomer according to claim 17 selected from the list consisting of amino acid residues 25-82 (SEQ ID NO. 5); amino acid residues 83-140 (SEQ ID NO. 6); amino acid residues 141-198 (SEQ ID NO. 7); amino acid residues 199-256 (SEQ ID NO. 8); amino acid residues 257-314 (SEQ ID NO. 9); and amino acid residues 315-368 (SEQ ID NO. 10); of the amino acid sequence set forth in Figure 1 (SEQ ID NO. 3).

19. A monomer according to claim 17 defined by the amino acid residues 1 to 24 (SEQ ID NO. 4) of the amino acid sequence set forth in Figure 1 (SEQ ID NO. 3).

20. A protease sensitive peptide comprising the amino acid sequence:



wherein X_1 and X_2 are preferably the same and are preferably both Lys residues and wherein R_1 and R_2 may be the same or different and each is a D or L amino acid, a peptide, a polypeptide, a protein, or an alkyl, substituted alkyl, alkenyl, substituted alkenyl, acyl, dienyl, arylalkyl, arylalkenyl, aryl, substituted aryl, heterocyclic, substituted heterocyclic, cycloalkyl, substituted cycloalkyl, halo, haloalkyl, nitro, hydroxy, thiol, sulfonyl, carboxy, alkoxy, aryloxy and alkylaryloxy group and the like.

21. A protease sensitive peptide according to claim 21 wherein R_1 and R_2 may be the same or different and each is a peptide or polypeptide and X_1 and X_2 are each Lys.

22. A protease sensitive peptide according to claim 20 or 21 in recombinant or synthetic form.

23. A nucleic acid molecule encoding the protease sensitive peptide according to claim 22.

24. A genetic construct comprising a nucleic acid molecule comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a type II serine PI precursor or monomer thereof from a plant wherein said precursor comprises at least three PI monomers and wherein at least one of said monomers has a chymotrypsin specific site and at least one of said other monomers has a trypsin specific site and wherein said genetic sequence further comprises expression means to permit expression of said nucleic acid molecule, replication means to permit replication in a plant cell or, alternatively, integration means, to permit stable integration of said nucleic acid molecule into a plant cell genome.

25. A transgenic plant carrying a genetic construct, said genetic construct comprising a deoxyribonucleic acid molecule which encodes a type II serine PI or monomer thereof, wherein said precursor comprises a sequence of nucleotides which encodes or is complementary to a sequence which encodes a type II serine proteinase inhibitor (PI) precursor from a plant wherein said precursor comprises at least three PI monomers and wherein at least one of said monomers has a chymotrypsin specific site and at least one other of said monomers has a trypsin specific site.

26. A transgenic plant according to claim 25 which produces one or more PI monomers selected from the listing consisting of amino acid residues 25-82 (SEQ ID NO. 5); amino acid residues 83-140 (SEQ ID NO. 6); amino acid residues 141-198 (SEQ ID NO. 7); amino acid residues 199-256 (SEQ ID NO. 8); amino acid residues 257-314 (SEQ ID NO. 9); and amino acid residues 315-368 (SEQ ID NO. 10) of the amino acid sequence set forth in Figure 1 (SEQ ID NO. 3).

27. A transgenic plant according to claim 25 which produces a PI monomer consisting of amino acid residues 1-24 (SEQ ID NO. 4) of the amino acid sequence set forth in SEQ ID NO. 3.

28. A method of increasing, enhancing or otherwise facilitating resistance of a plant to insect or other pathogen infestation, said method comprising introducing a nucleic acid molecule as defined in claim 1 or 7 or 10 or 11 into a cell or group of cells of said plant, regenerating a plant therefrom and growing said plant for a time and under conditions sufficient to permit expression of said nucleic acid molecule into a PI or precursor thereof capable of inhibiting growth and/or infestation by said pathogen.

1 / 20

2/20
3 / 20
4/20
5 / 20
6/20

FIG 1

2/20

18

CGA GTA AGT ATG GCT GTT
Arg Val Ser Met Ala Val

-29

66

CAC AGA GTT AGT TTC CTT GCT CTC CTC TTA TTT GGA ATG TCT CTG
His Arg Val Ser Phe Leu Ala Leu Leu Leu Phe Gly Met Ser Leu

-20

114

CTT GTA AGC AAT GTG GAA CAT GCA GAT GCC AAG GCT TGT ACC TTA AAC
Leu Val Ser Asn Val Glu His Ala Asp Ala Lys Ala Cys Thr Leu Asn
-10 -1 1 5

162

TGT GAT CCA AGA ATT GCC TAT GGA GTT TGC CCG CGT TCA GAA GAA AAG
Cys Asp Pro Arg Ile Ala Tyr Gly Val Cys Pro Arg Ser Glu Glu Lys
10 15 20

210

AAG AAT GAT CGG ATA TGC ACC AAC TGT TGC GCA GGC ACG AAG GGT TGT
Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Thr Lys Gly Cys
25 30 35

258

AAG TAC TTC AGT GAT GAT GGA ACT TTT GTT TGT GAA GGA GAG TCT GAT
Lys Tyr Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp
40 45 50

FIG 1

3/20

CCT AGA AAT CCA AAG GCT TGT ACC TTA AAC TGT GAT CCA AGA ATT GCC	306
Pro Arg Asn Pro Lys Ala Cys Thr Leu Asn Cys Asp Pro Arg Ile Ala	70
55	60
	65
TAT GGA GTT TGC CCG CGT TCA GAA AAG AAG AAT GAT CGG ATA TGC	354
Tyr Gly Val Cys Pro Arg Ser Glu Glu Lys Lys Asn Asp Arg Ile Cys	80
	75
	85
ACC AAC TGT TGC GCA GGC ACG AAG GGT TGT AAG TAC TTC AGT GAT GAT	402
Thr Asn Cys Cys Ala Gly Thr Lys Gly Cys Lys Tyr Phe Ser Asp Asp	100
	95
GGA ACT TTT GTT TGT GAA GAG TCT GAT CCT AGA AAT CCA AAG GCT	450
Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Arg Asn Pro Lys Ala	110
	105
	115
TGT CCT CGG AAT TGC GAT CCA AGA ATT GCC TAT GGG ATT TGC CCA CTT	498
Cys Pro Arg Asn Cys Asp Pro Arg Ile Ala Tyr Gly Ile Cys Pro Leu	120
	125
	130
GCA GAA GAA AAG AAG AAT GAT CGG ATA TGC ACC AAC TGT TGC GCA GGC	546
Ala Glu Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly	140
	145
	150

FIG 1

4/20

AAA AAG GGT TGT AAG TAC TTT AGT GAT GAT GGA ACT TTT GTT TGT GAA 594
Lys Lys Gly Cys 155
 160
 165
 GGA GAG TCT GAT CCT AAA AAT CCA AAG GCC TGT CCT CGG AAT TGT GAT 642
 Gly Glu Ser Asp Pro Lys Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp
 170 175 180
 GGA AGA ATT GCC TAT GGG ATT TGC CCA CTT TCA GAA GAA AAG AAG AAT 690
 Gly Arg Ile Ala Tyr Gly Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn
 185 190 195
 GAT CGG ATA TGC ACC AAC TGC TGC GCA GGC AAA AAG GGT TGT AAG TAC 738
 Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr
 200 205 210
 TTT AGT GAT GAT GGA ACT TTT GTT TGT GAA GGA GAG TCT GAT CCT AAA 786
 Phe Ser Asp Asp Gly Thr Phe Val Cys Glu Gly Glu Ser Asp Pro Lys
 215 220 225 230
 AAT CCA AAG GCT TGT CCT CGG AAT TGT GAT GAT GGA AGA ATT GCC TAT GGG 834
 Asn Pro Lys Ala Cys Pro Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly
 235 240 245

FIG 1

5/20

882	ATT TGC CCA CTT TCA GAA GAA AAG AAG AAT GAT CGG ATA TGC ACA AAC	
	Ile Cys Pro Leu Ser Glu Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn	250 255 260
930	TGT TGC GCA GGC AAA AAG AAG GGC TGT AAG TAC TTT AGT GAT GAT GGA ACT	
	Cys Cys Ala Gly Lys Lys Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr	265 270 275
978	TTT GTT TGT GAA GGA GAG GAG TCT GAT CCT ACA AAT CCA AAG GCC TGT CCT	
	Phe Val Cys Glu Gly Glu Ser Asp Pro Arg Asn Pro Lys Ala Cys Pro	280 285 290
1026	CGG AAT TGT GAT GGA AGA ATT GCC TAT GGA ATT TGC CCA CTT TCA GAA	
	Arg Asn Cys Asp Gly Arg Ile Ala Tyr Gly Ile Cys Pro Leu Ser Glu	295 300 305 310
1074	GAA AAG AAG AAT GAT CGG ATA TGC ACC AAT TGT TGC GCA GGC AAG AAG	
	Glu Lys Lys Asn Asp Arg Ile Cys Thr Asn Cys Cys Ala Gly Lys Lys	315 320 325
1122	GGC TGT AAG TAC TTT ACT GAT GAT GGA ACT TTT ATT TGT GAA GGA GAA	
	Gly Cys Lys Tyr Phe Ser Asp Asp Gly Thr Phe Ile Cys Glu Gly Glu	330 335 340

FIG 1

6/20

1170
TCT GAA TAT GCC AGC AAA GTG GAT GAA TAT GTT GGT GAA GTG GAG AAT
Ser Glu Tyr Ala Ser Lys Lys Val Asp Glu Tyr Val Gly Glu Val Glu Asn
345 350 355

1220
GAT CTC CAG AAG TCT AAG GTT GCT GCT TCC TAAGTCCTAA CTAATAATAT
Asp Leu Gln Lys Ser Lys Val Ala Val Ser
360 365

1280
GTAGTCTATG TATGAAACAA AGGCATGCCA ATATGCTCTG TCTTGCCCTGT AATCTGTAAT

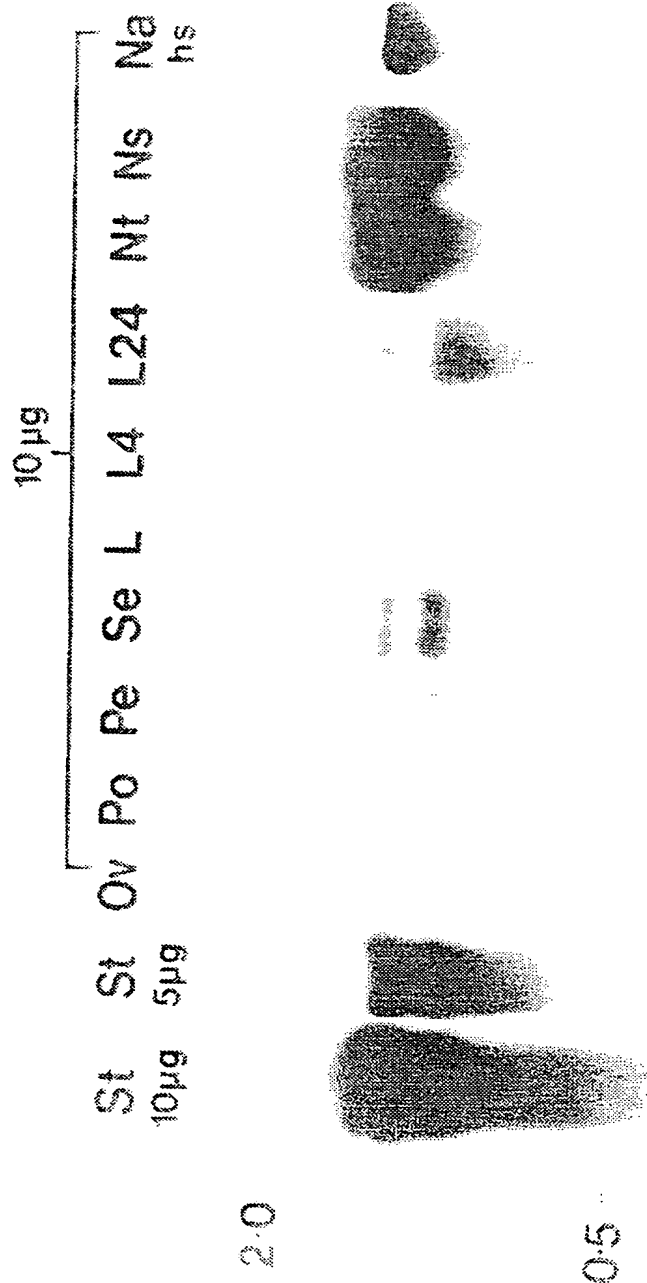
1340
ATGGTAGTGG AGCTTTTCCA CTGCCCTGTT AATAAGAAAT GGAGCACTAG TTTGTTTTAG

1360
TTAAAAAAA AAAAAAAA

FIG 1

7 / 20

FIG 2



8/20

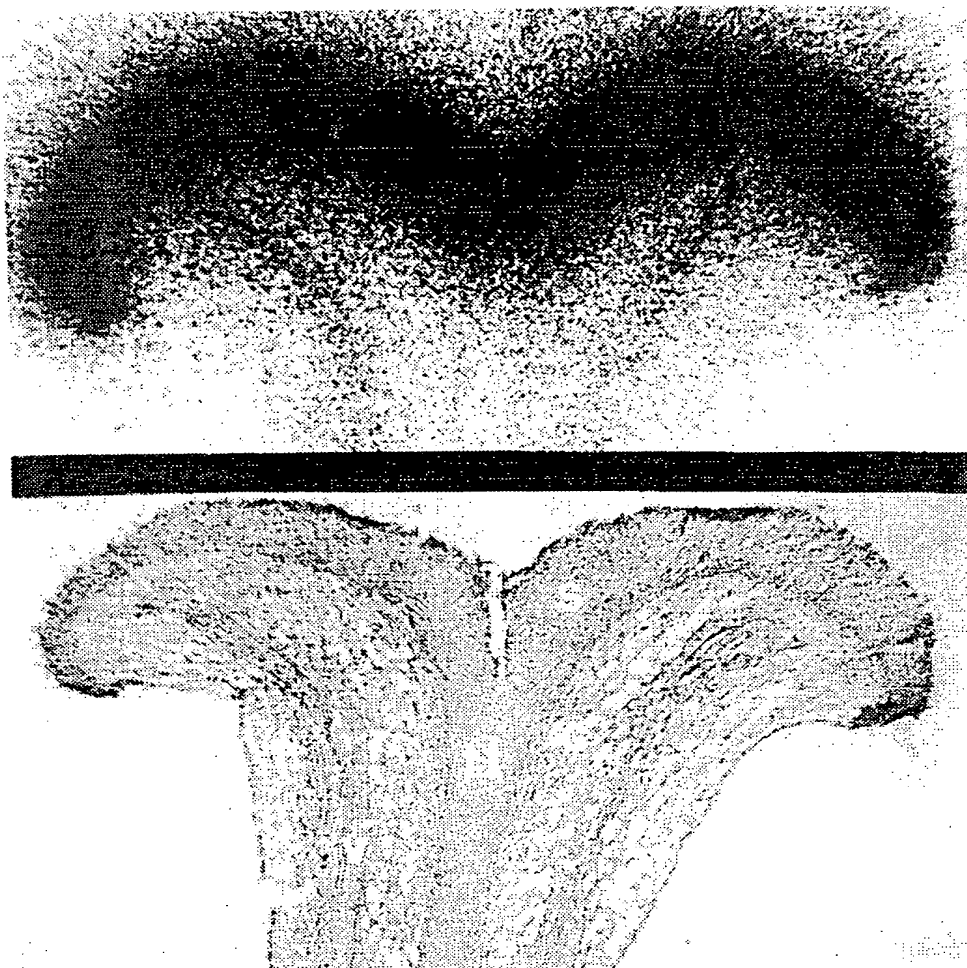


FIG 3

9/20

EcoRI HindIII

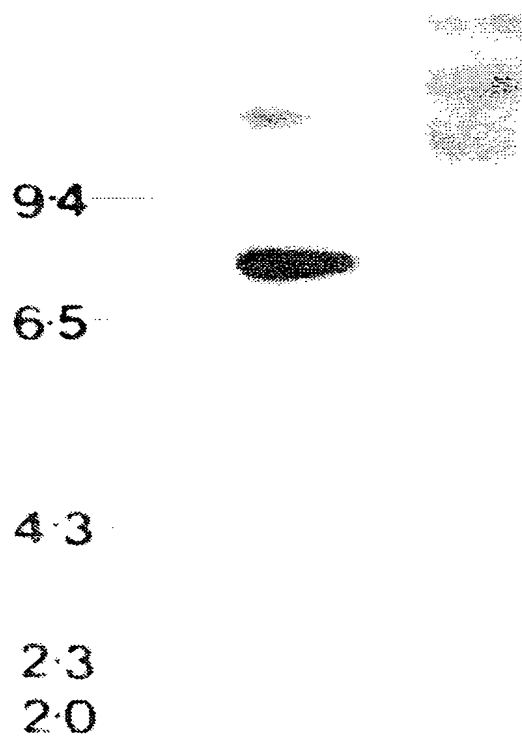
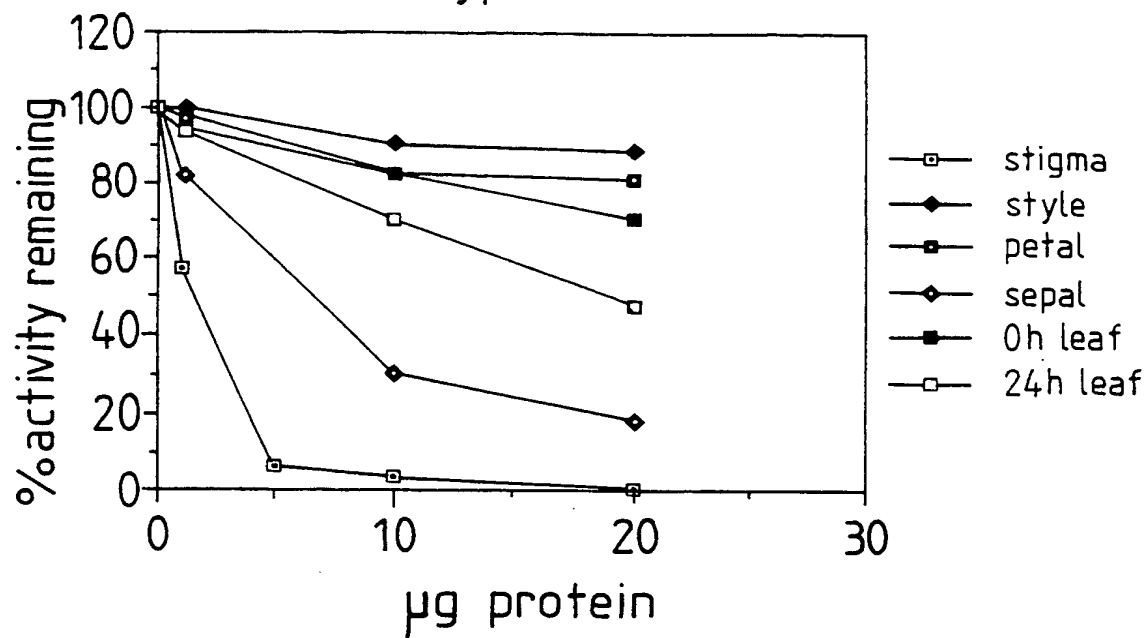


FIG 4

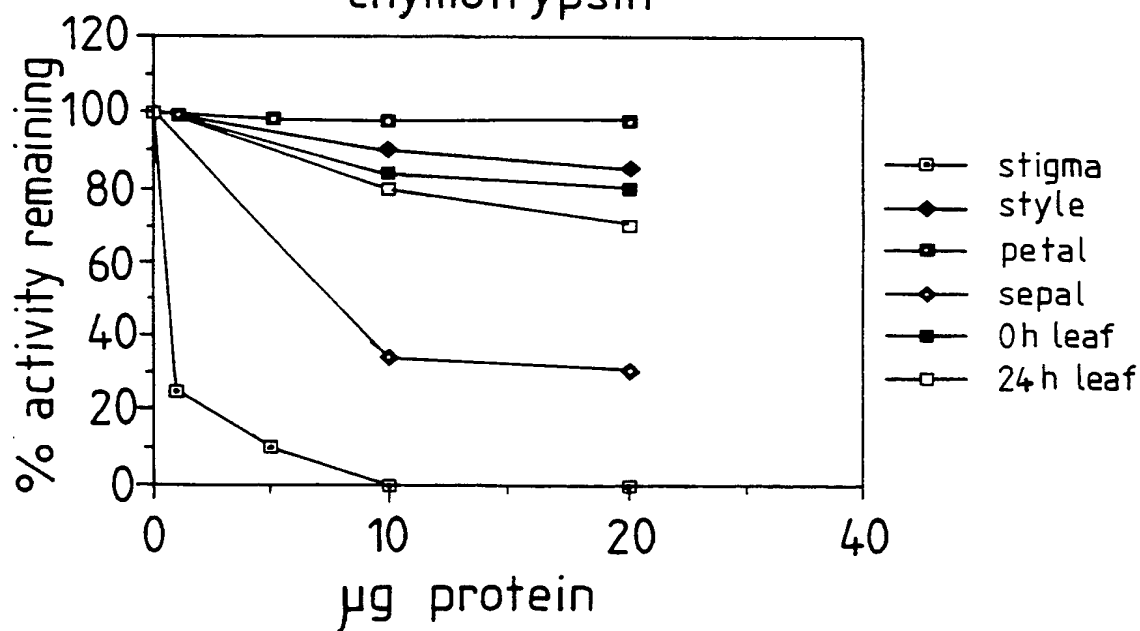
10/ 20

FIGURE 5A

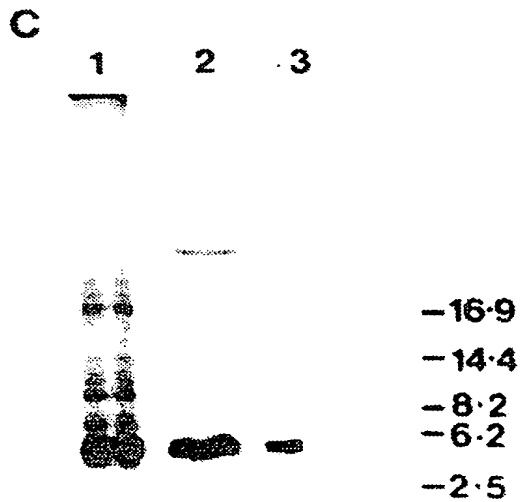
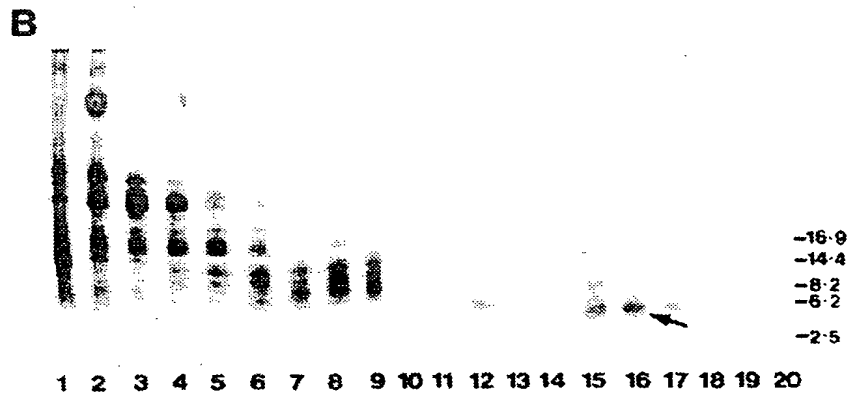
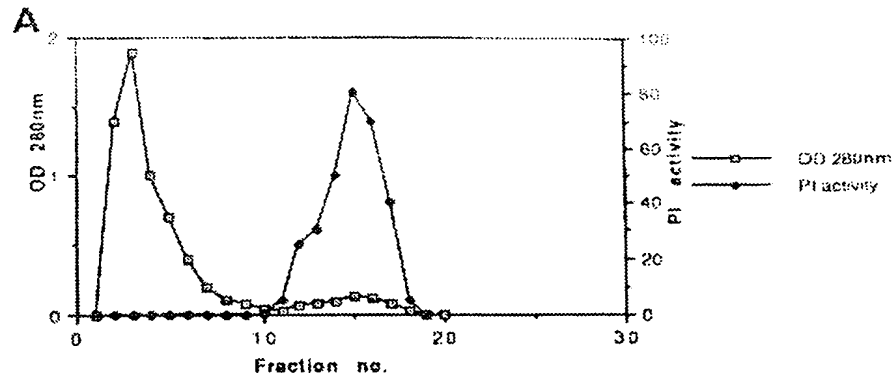
trypsin

FIGURE 5B

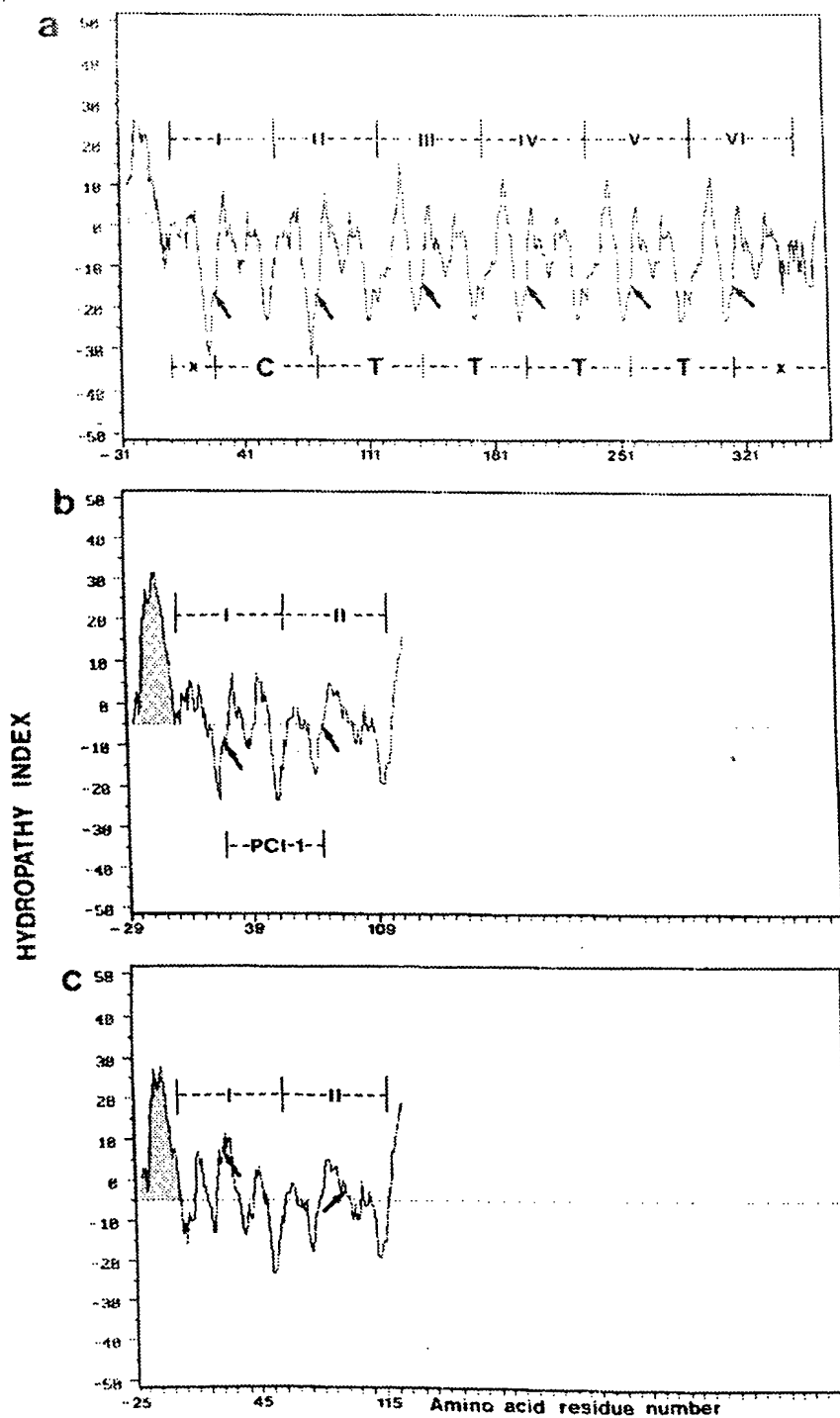
chymotrypsin



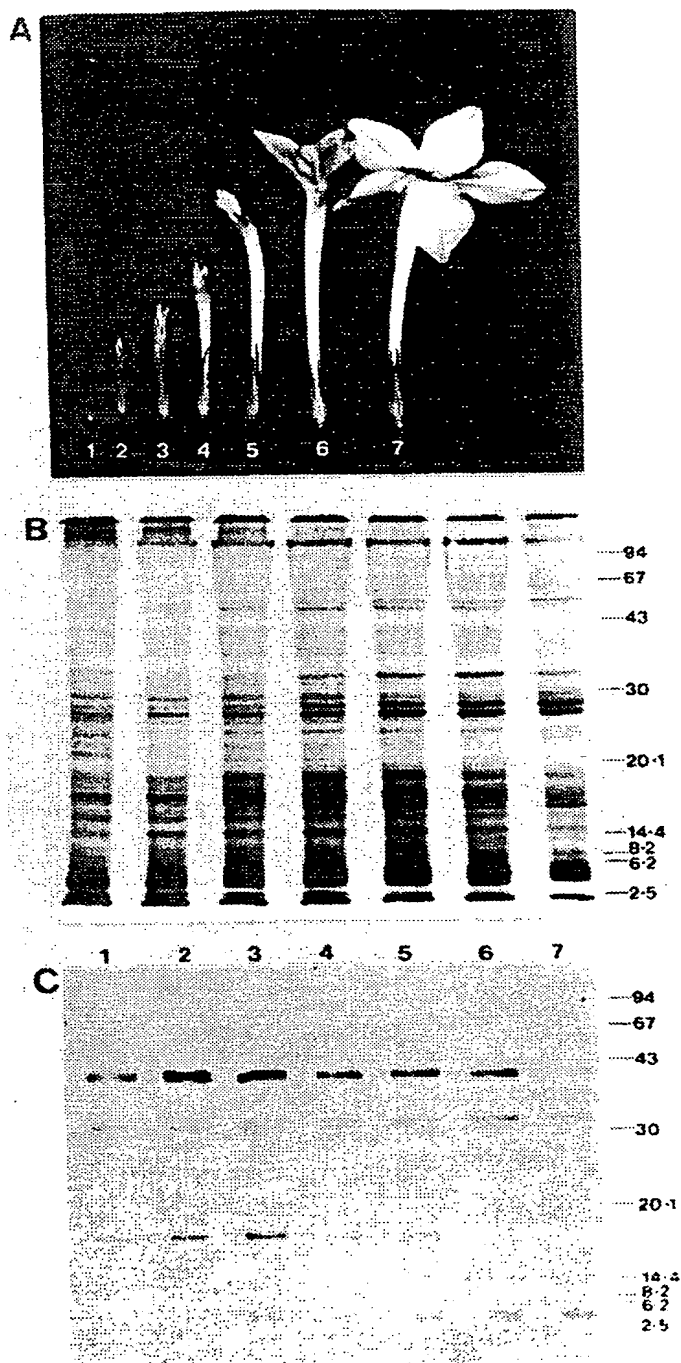
11/20

FIG 6

12/20

FIG 7

13/20

FIG 8

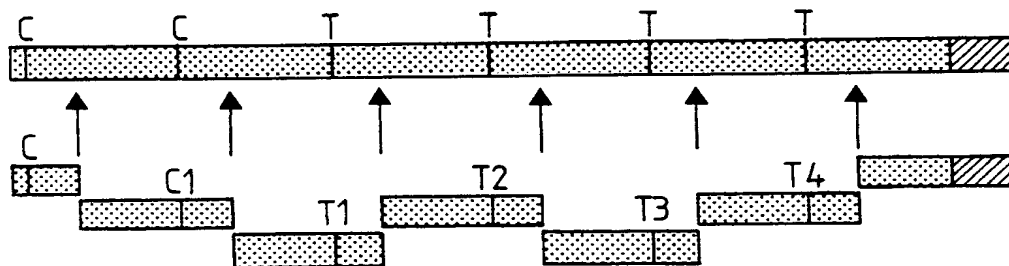
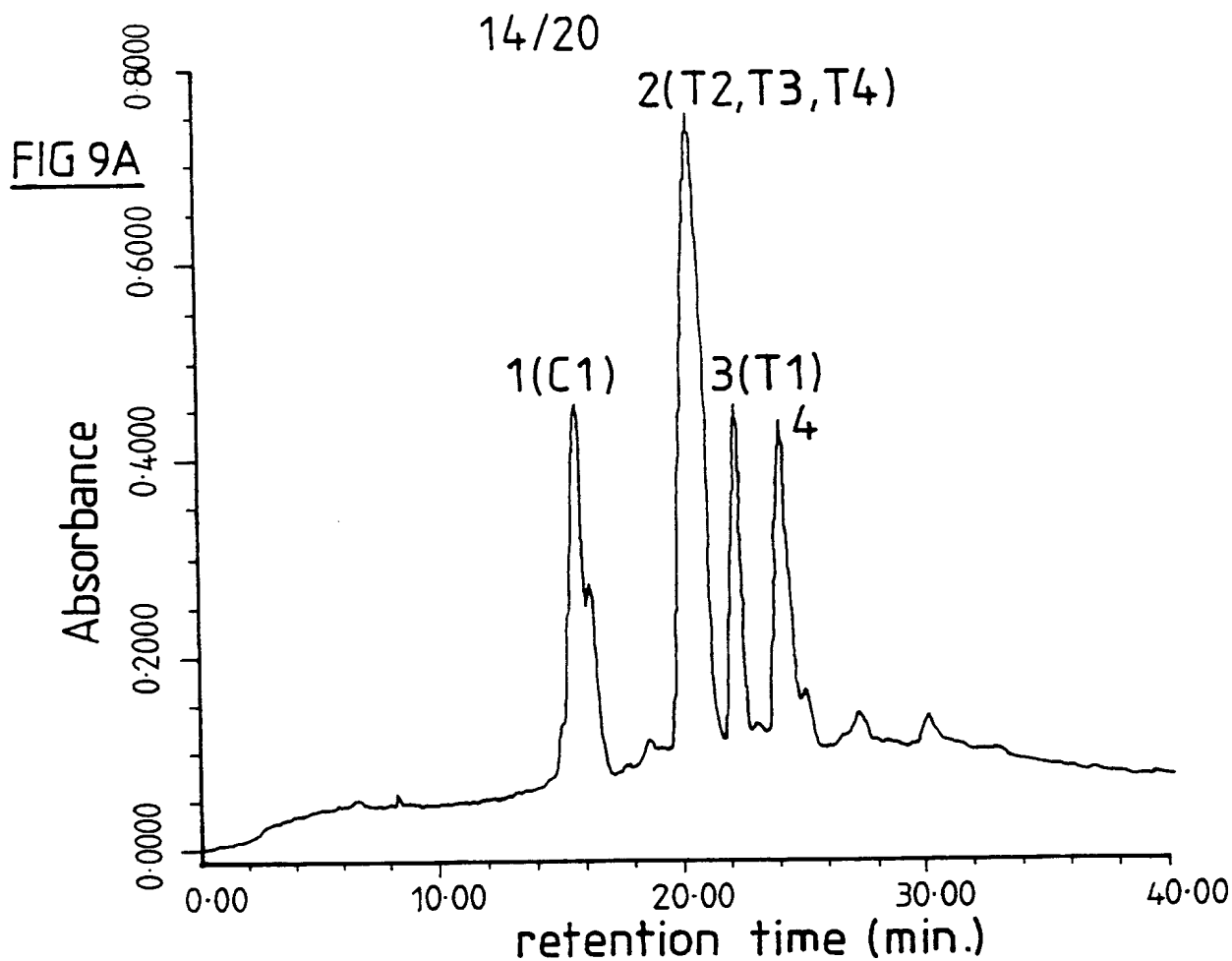


FIG 9B

C1 DRICTNCCAGTKGCKYFSDDGTFVCEGESDPRNPKACTTLNCDPRIAYGVCPRS
 T1 DRICTNCCAGTKGCKYFSDDGTFVCEGESDPRNPKACPRNCDPRIAYGICPL
 T2 DRICTNCCAGKKKGCKYFSDDGTFVCEGESDPKNPKACPRNCDGRIAYGICPLS
 T3 DRICTNCCAGKKKGCKYFSDDGTFVCEGESDPKNPKACPRNCDGRIAYGICPLS
 T4 DRICTNCCAGKKKGCKYFSDDGTFVCEGESDPRNPKACPRNCDGRIAYGICPLS

1 10 20 30 40 50

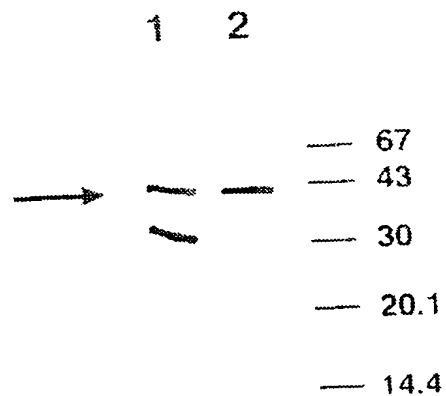
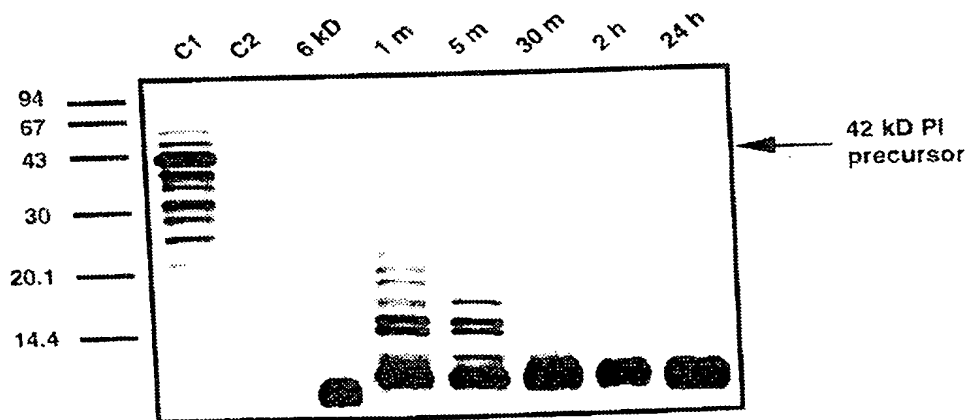
FIGURE 9C

15/20

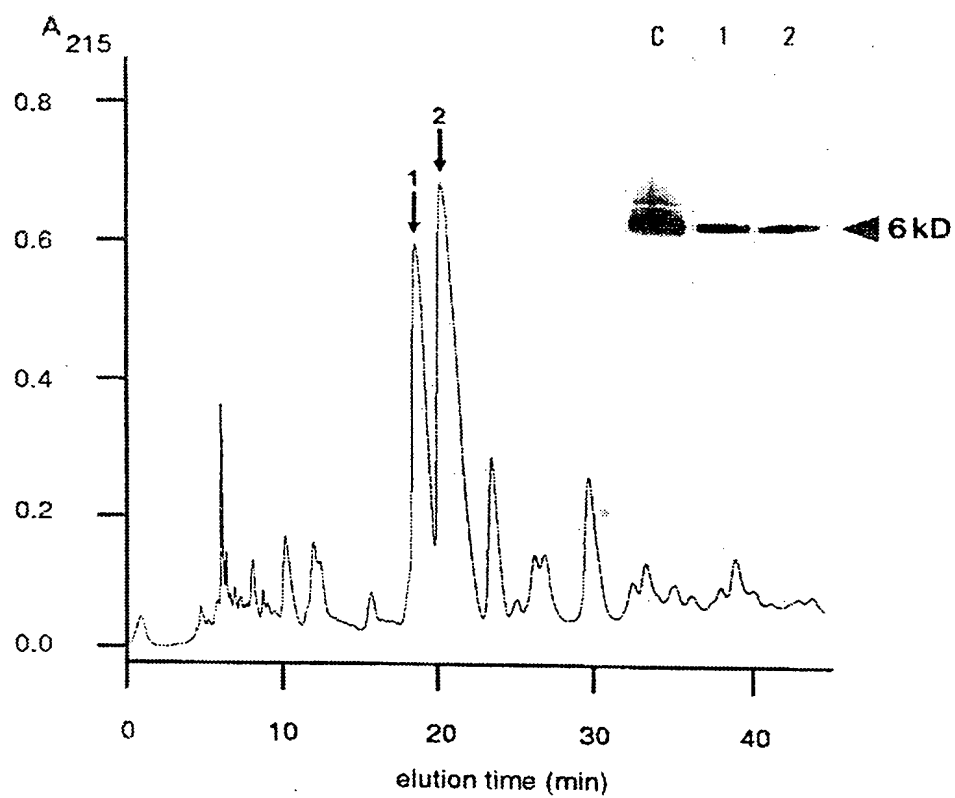
-10
1 10
ICP(R or L)(S or A)EEKKNDRICTNCCAG(T or K)KG

FIGURE 10

16/20

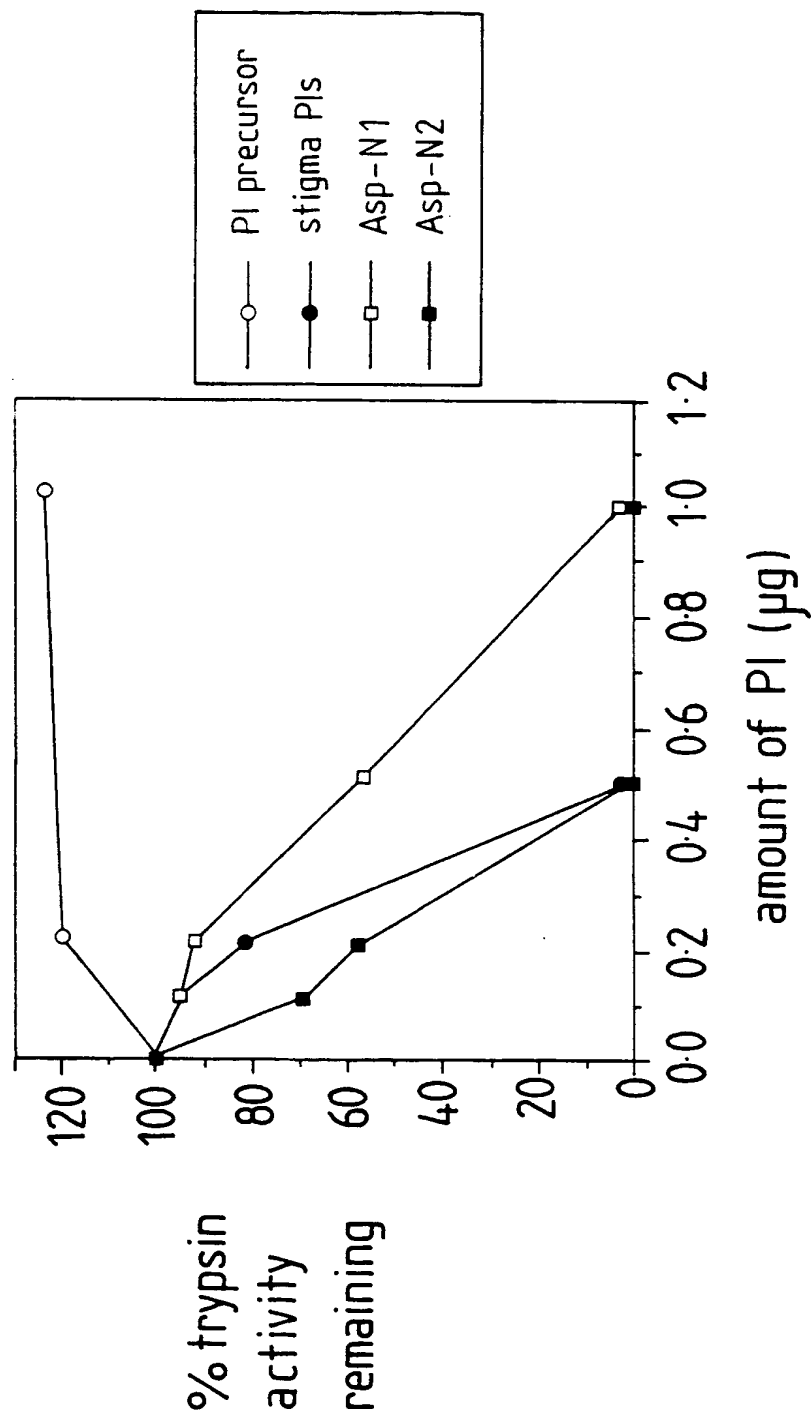
FIG 11AFIG 11B

17 / 20

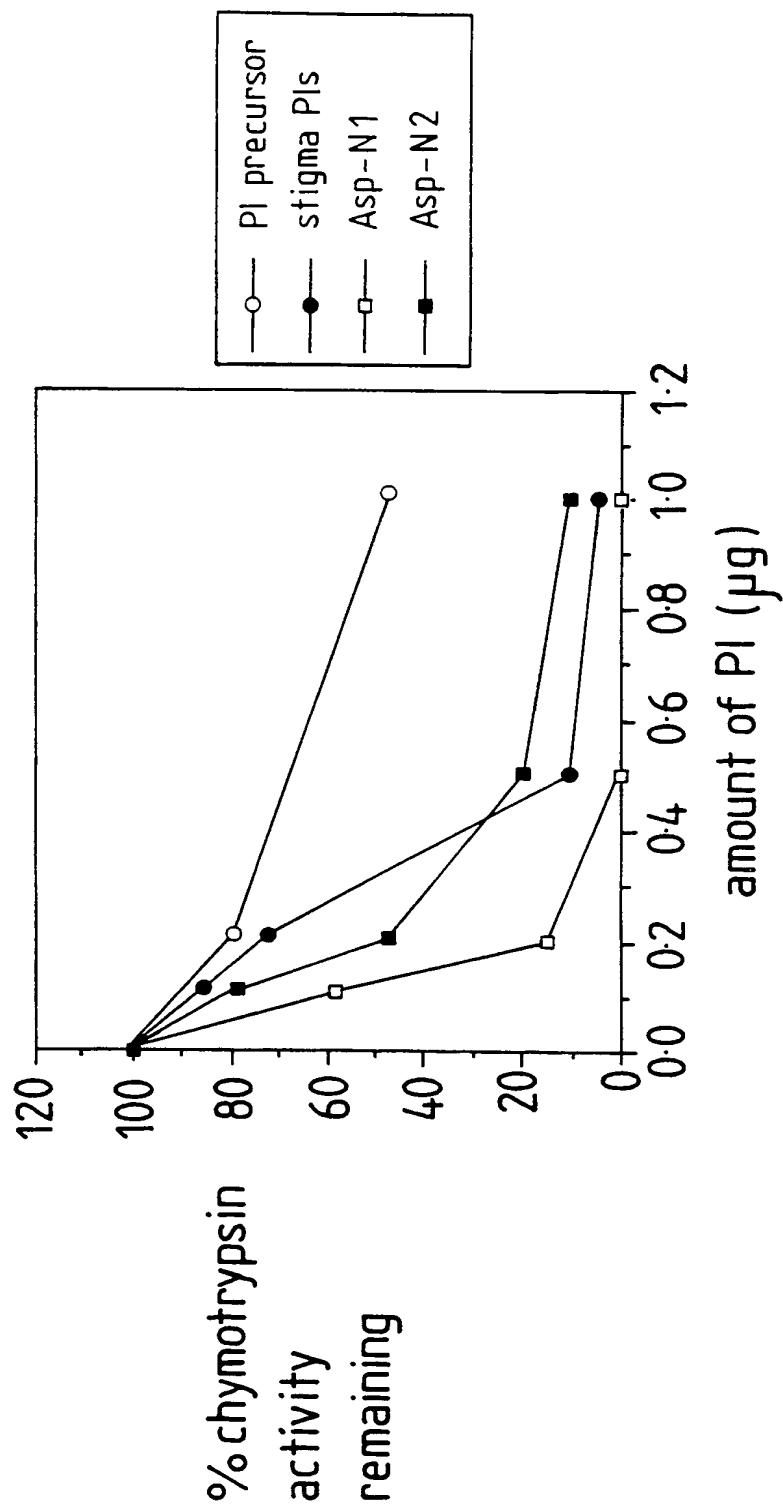
FIG 12

18/20

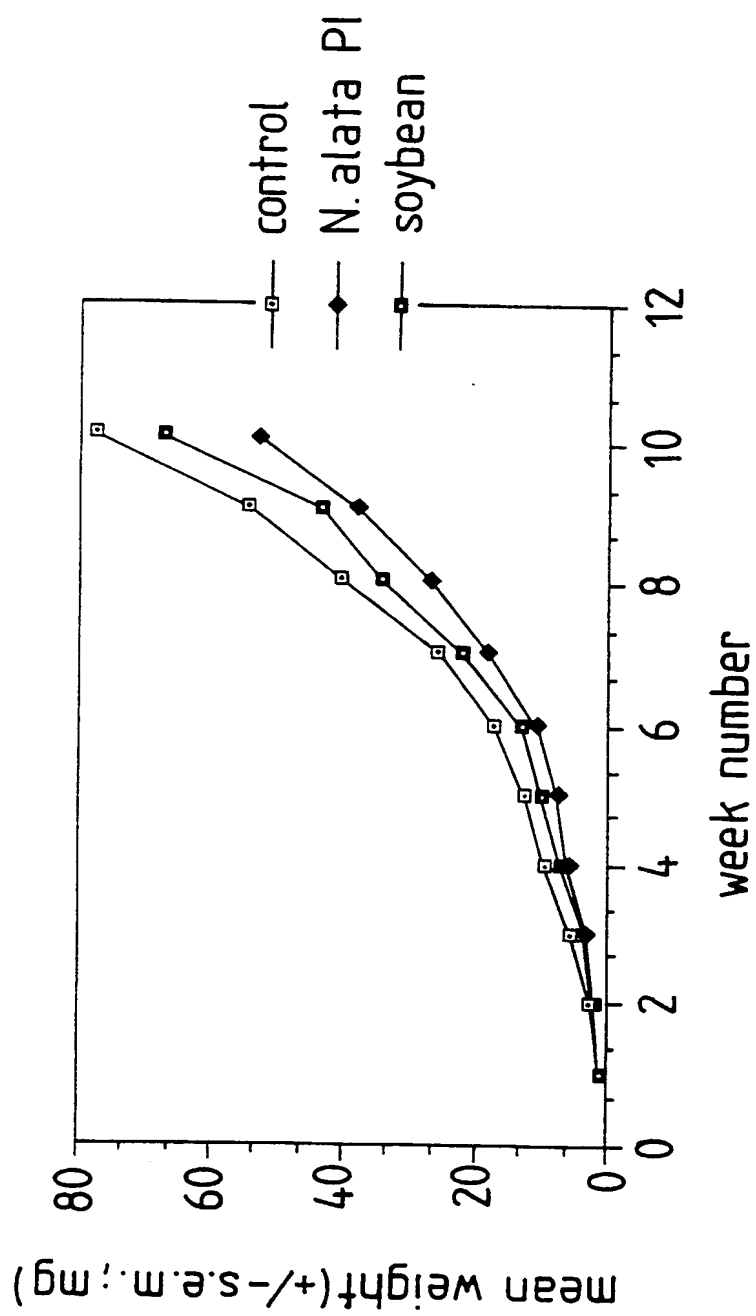
FIGURE 13A



19 / 20

FIGURE 13B

20 / 20

FIGURE 14

A. CLASSIFICATION OF SUBJECT MATTER Int. Cl. ⁵ C12N 15/29; A01H 1/00; C07K 5/10, 7/06, 7/08, 7/10, 13/00 According to International Patent Classification (IPC) or to both national classification and IPC					
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC: C12N; Derwent database WPAT, ACS database CASM, Keywords as below Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched IPC: AU database C12N 15/29 Electronic data base consulted during the international search (name of data base, and where practicable, search terms used) CASM, WPAT, Biotechnology Abstracts (BIOT-Derwent database), Keywords: serine, prote:, peptidase#, chymotrypsin, trypsin, inhibit:, plant#					
C. DOCUMENTS CONSIDERED TO BE RELEVANT					
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.			
A	Biochemistry, Vol. 15 (No. 16), issued 1976, Bryant, J. et al, "Proteinase inhibitor II from potatoes: Isolation and characterization of its protomer components", pages 3418-3423				
A	Archives of Biochemistry and Biophysics, Vol. 213 (No. 2), issued 1982, Plunkett, G. et al, "Proteinase inhibitors I and II from leaves of wounded tomato plants: Purification and properties", pages 463-472				
<div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <input type="checkbox"/> Further documents are listed in the continuation of Box C. </div> <div style="width: 45%;"> <input type="checkbox"/> See patent family annex. </div> </div>					
<table style="width: 100%; border: none;"> <tr> <td style="width: 30%; vertical-align: top;"> * Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed </td> <td style="width: 10%; vertical-align: top; padding-left: 20px;"> "T" "X" "Y" "&" </td> <td style="width: 60%; vertical-align: top; padding-left: 20px;"> later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art document member of the same patent family </td> </tr> </table>			* Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed	"T" "X" "Y" "&"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art document member of the same patent family
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Date of the actual completion of the international search 30 March 1994 (30.03.94)		Date of mailing of the international search report <div style="font-size: 1.2em; font-family: cursive;">7 April 1994 (7.4.94)</div>			
Name and mailing address of the ISA/AU AUSTRALIAN INDUSTRIAL PROPERTY ORGANISATION PO BOX 200 WODEN ACT 2606 AUSTRALIA Facsimile No. 06 2853929		Authorized officer <div style="font-size: 1.2em; font-family: cursive;">Ian Rose</div> IAN ROSE Telephone No. (06) 2832494			

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international search report has not established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claim Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Claims 1-19, 24-28 directed to a type II serine proteinase inhibitor from plants, nucleic acid sequences coding therefore and transgenic plants which express the gene.

Claims 20-23 directed to a protease sensitive peptide which comprises a specific amine acid sequence which is sensitive to cleavage by proteases.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-19, 24-28

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

